

**Gateway[®]-Technologie:
Potenzial und Anwendungen in der
molekularen Pflanzenforschung**

Andrea Kolpack

Heike Loschelder





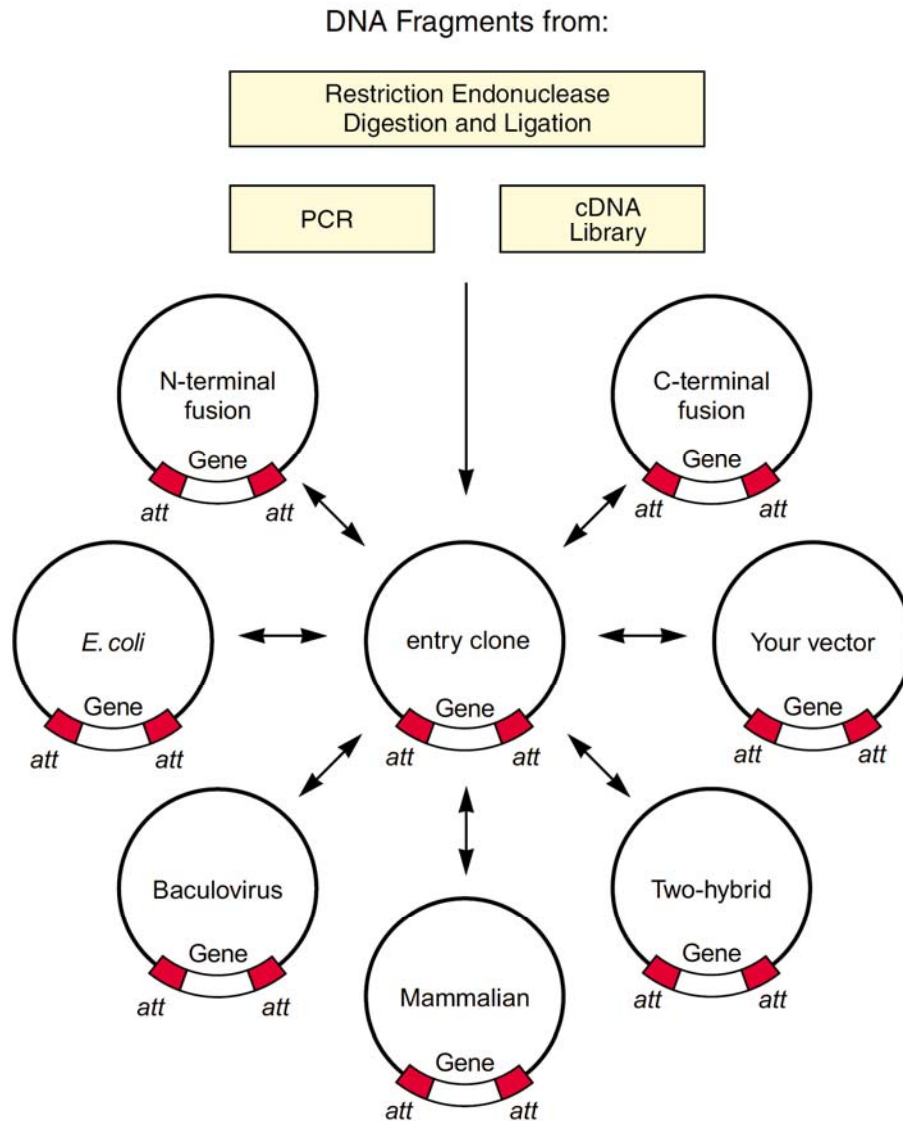
1. Overview and Entry Options
2. Destination Vectors

Key Benefits of the Gateway[®] Technology

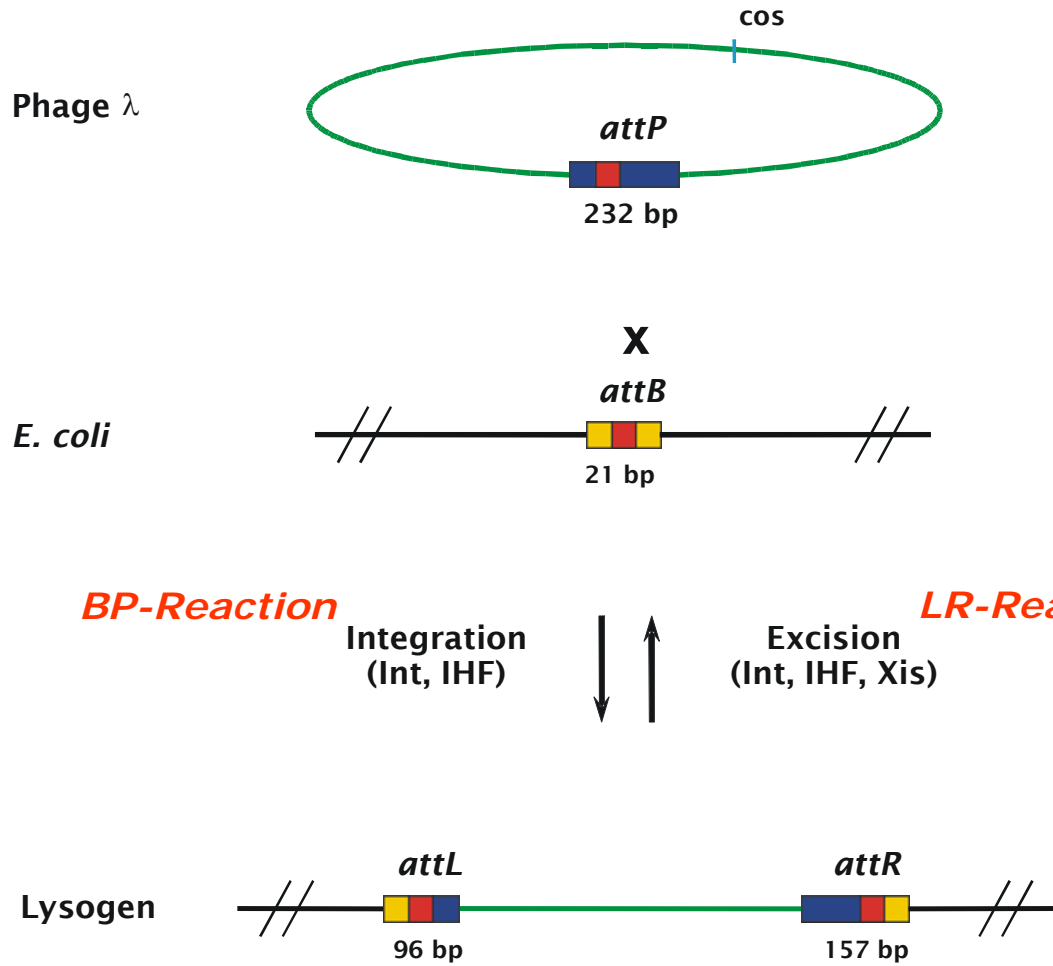


- Efficiently and easily shuttle insert DNA from one expression plasmid to another
- Simplify the cloning workflow and save time
- Create expression clones without using restriction enzymes and ligase
- Utilize ORF clones, a pre-made Gateway[®] collection
- Simultaneously clone, in a specific order and orientation, up to 4 DNA fragments into one plasmid

Gateway® Technology : Overview



Phage lambda recombination in *E. coli*



The Gateway® System
relies on five sets of
specific and non cross-
reacting *att* sequences

Characteristics of the Modified *att* Sites



Site	Length	Found in...
<i>attB</i>	25 bp	Expression vector Expression clone
<i>attP</i>	200 bp	Donor vector
<i>attL</i>	100 bp	Entry vector Entry clone
<i>attR</i>	125 bp	Destination vector

Specificity:

- *attB1* sites react only with *attP1* sites
- *attB2* sites react only with *attP2* sites
- *attL1* sites react only with *attR1* sites
- *attL2* sites react only with *attR2* sites

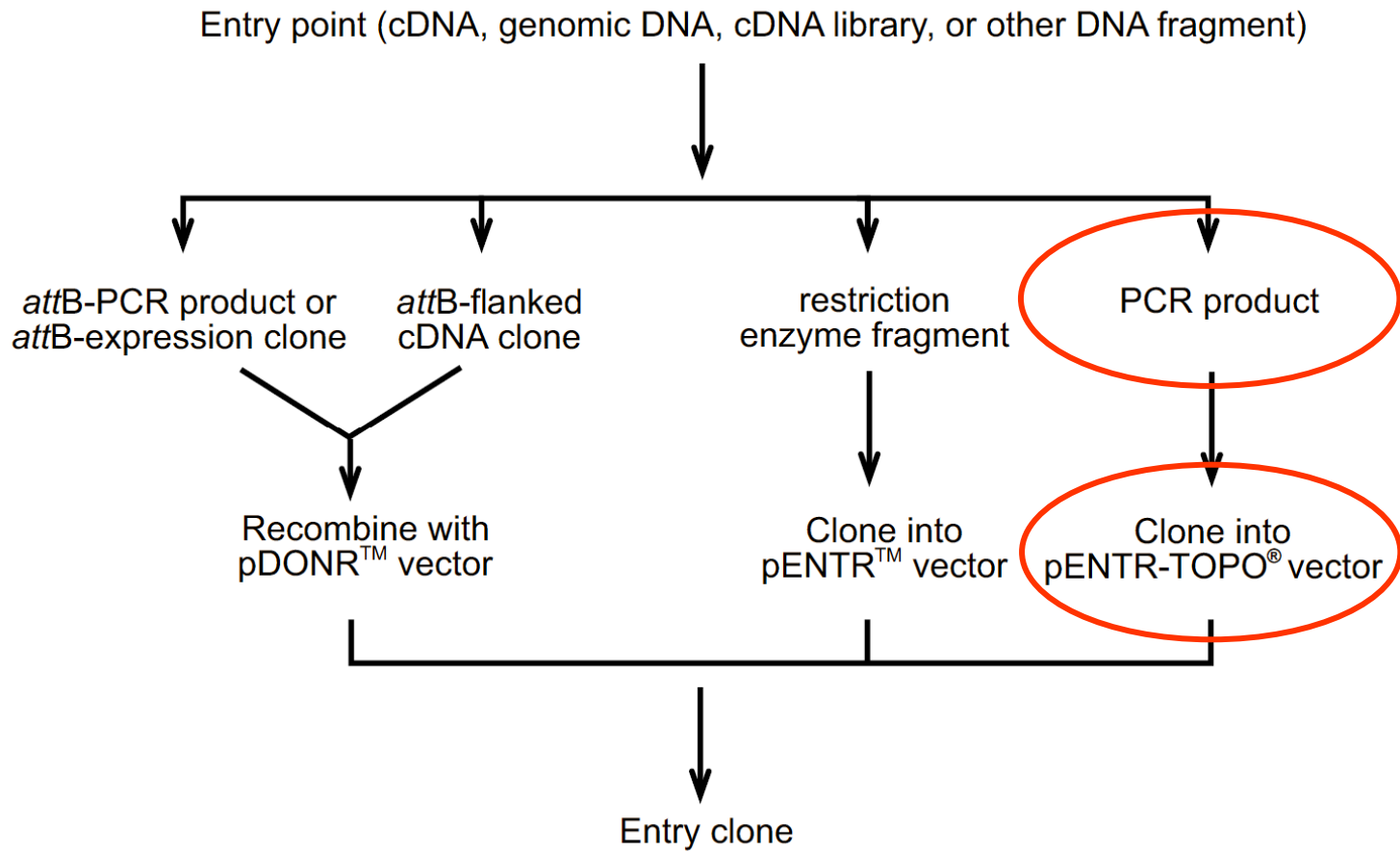
Options to Create Entry Clones



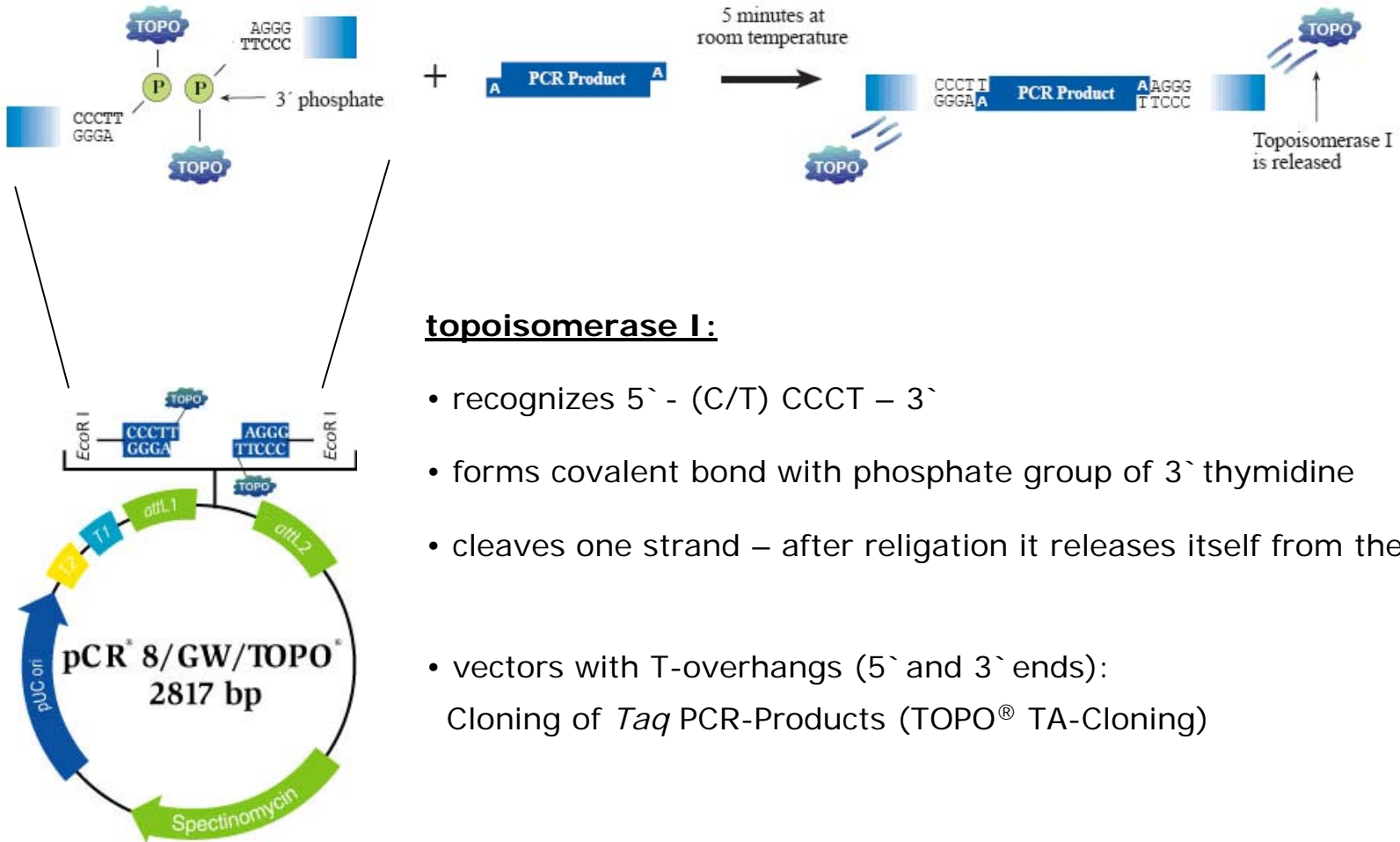
Entry Vector	Kozak	Shine-Dalgarno
pENTR/D-TOPO [®]	•	
pENTR/SD/D-TOPO [®]	•	•
pENTR [™] 1A	•	•
pENTR [™] 2B	•	
pENTR [™] 3C	•	•
pENTR [™] 4	•	
pENTR [™] 11	•	•

Vector	M13 Sequencing Sites	Selection Marker
pDONR [™] 201	No	Kanamycin
pDONR [™] 221	Yes	Kanamycin
pDONR [™] /Zeo	Yes	Zeocin [™]

Options to Create Entry Clones I



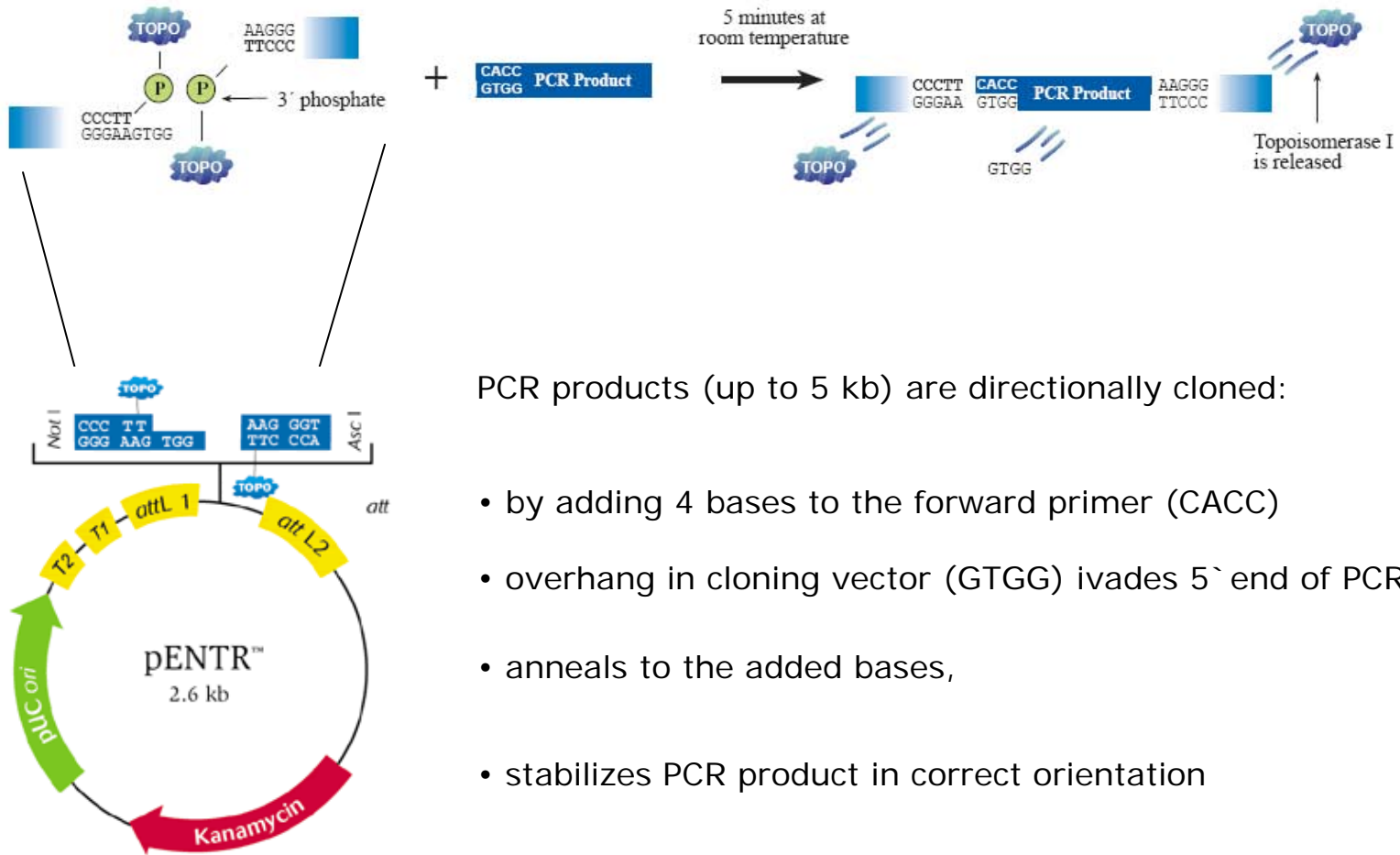
TOPO[®] - Cloning: TOPO[®]TA



topoisomerase I:

- recognizes 5` - (C/T) CCCT – 3`
- forms covalent bond with phosphate group of 3` thymidine
- cleaves one strand – after religation it releases itself from the DNA
- vectors with T-overhangs (5` and 3` ends):
Cloning of *Taq* PCR-Products (TOPO[®] TA-Cloning)

PCR-Directional TOPO® Cloning



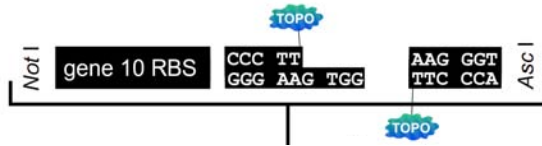
PCR products (up to 5 kb) are directionally cloned:

- by adding 4 bases to the forward primer (CACC)
- overhang in cloning vector (GTGG) invades 5' end of PCR-product,
- anneals to the added bases,
- stabilizes PCR product in correct orientation

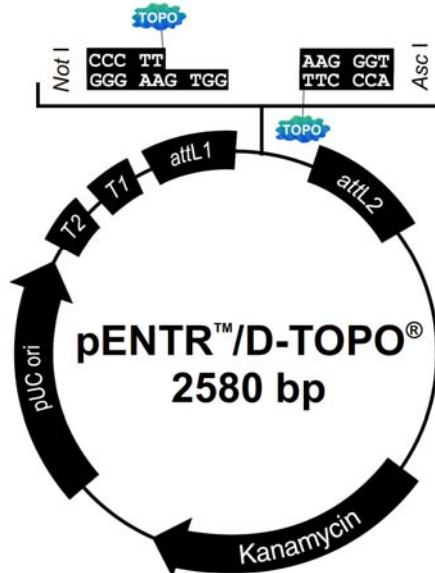
PCR-Directional TOPO[®] Cloning

A

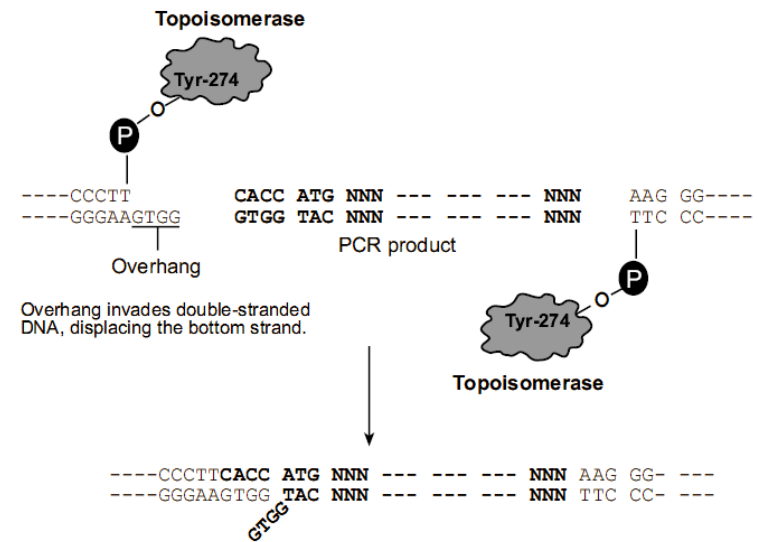
pENTR/SD/D-TOPO



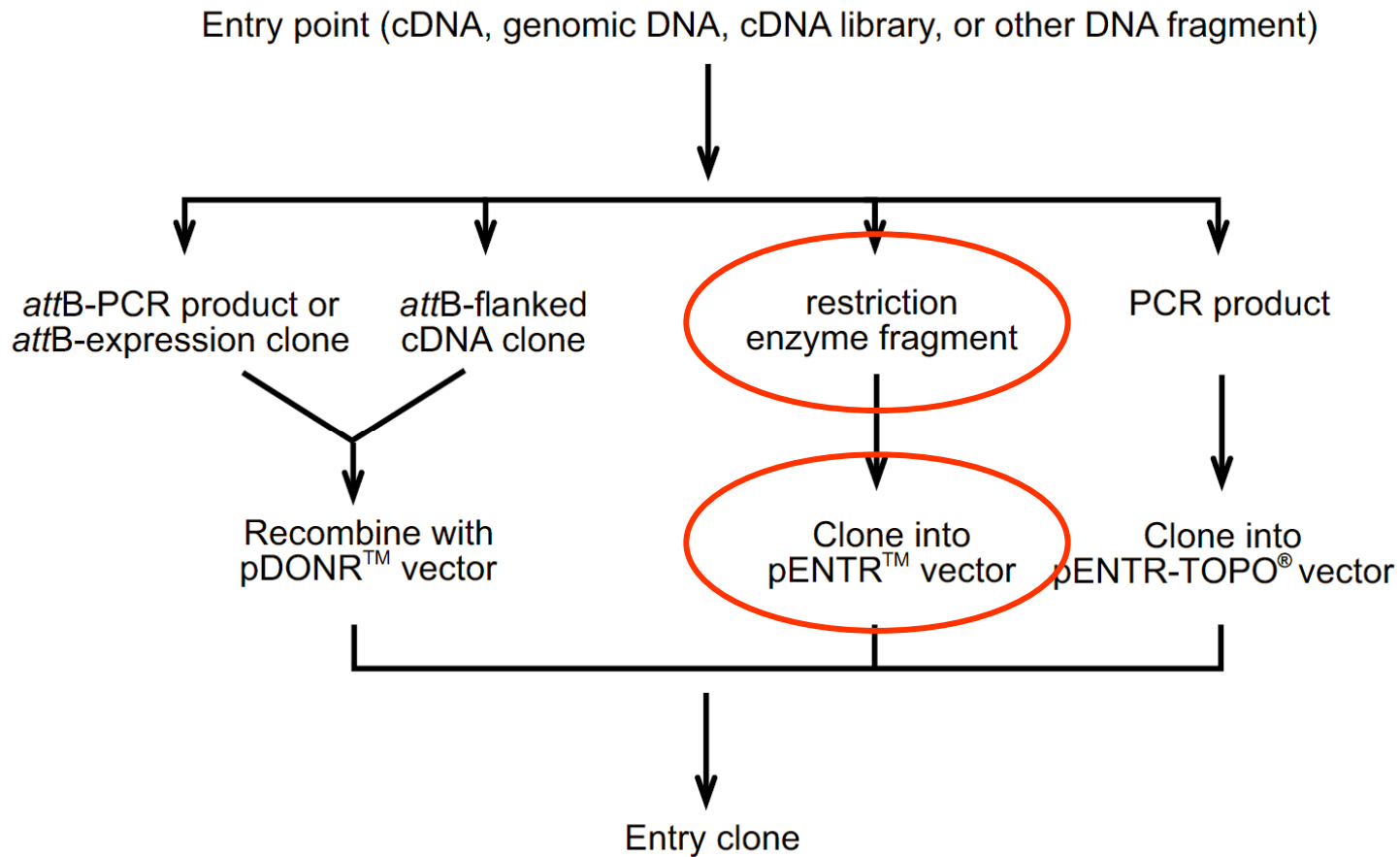
pENTR/D/D-TOPO



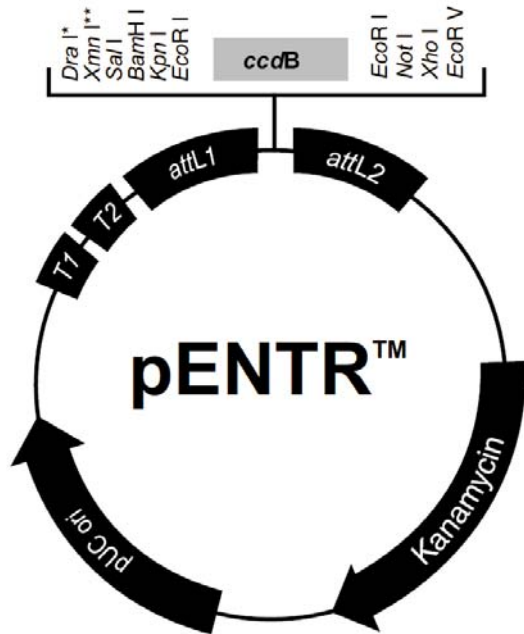
B



Options to Create Entry Clones II



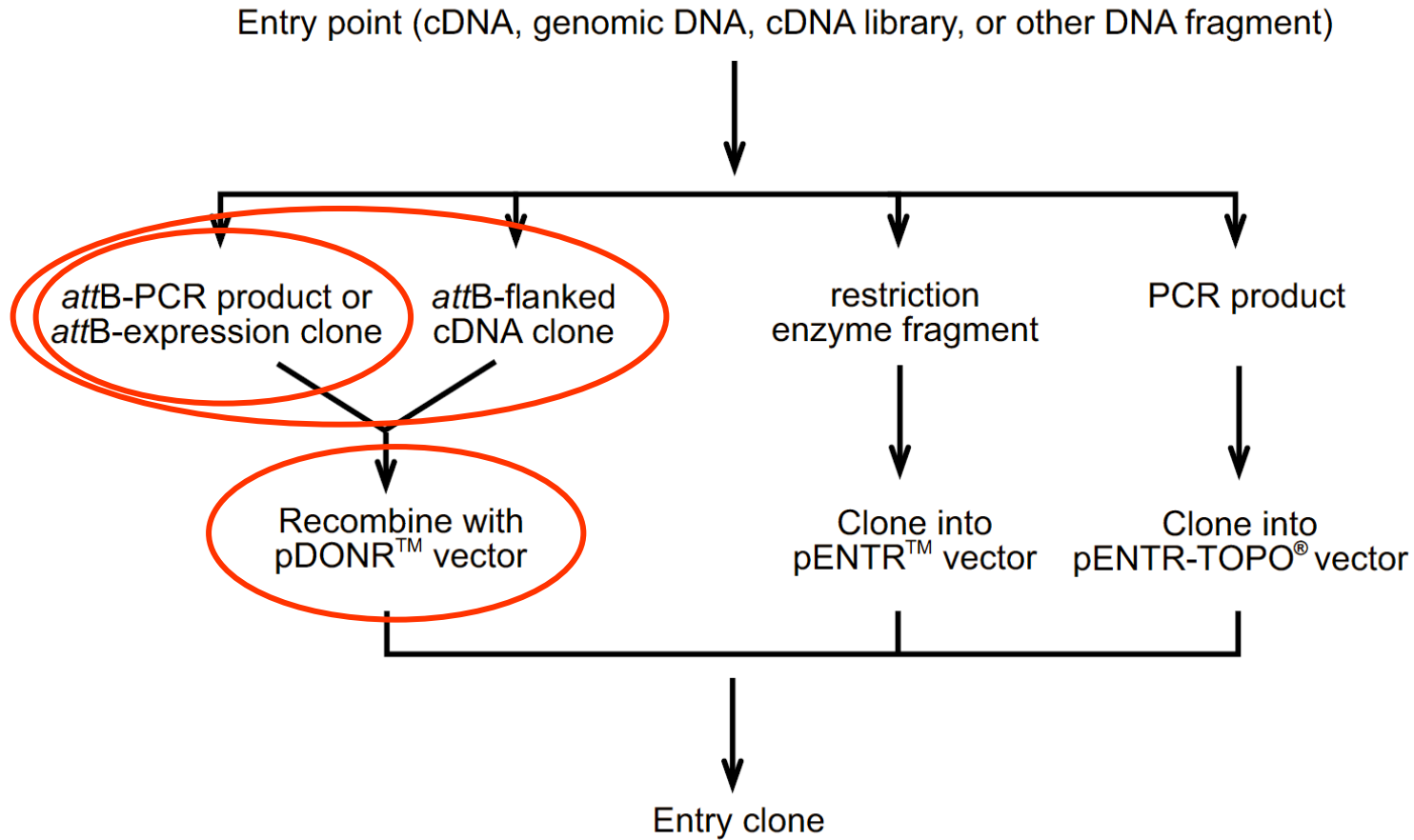
Restriction/Ligase Cloning



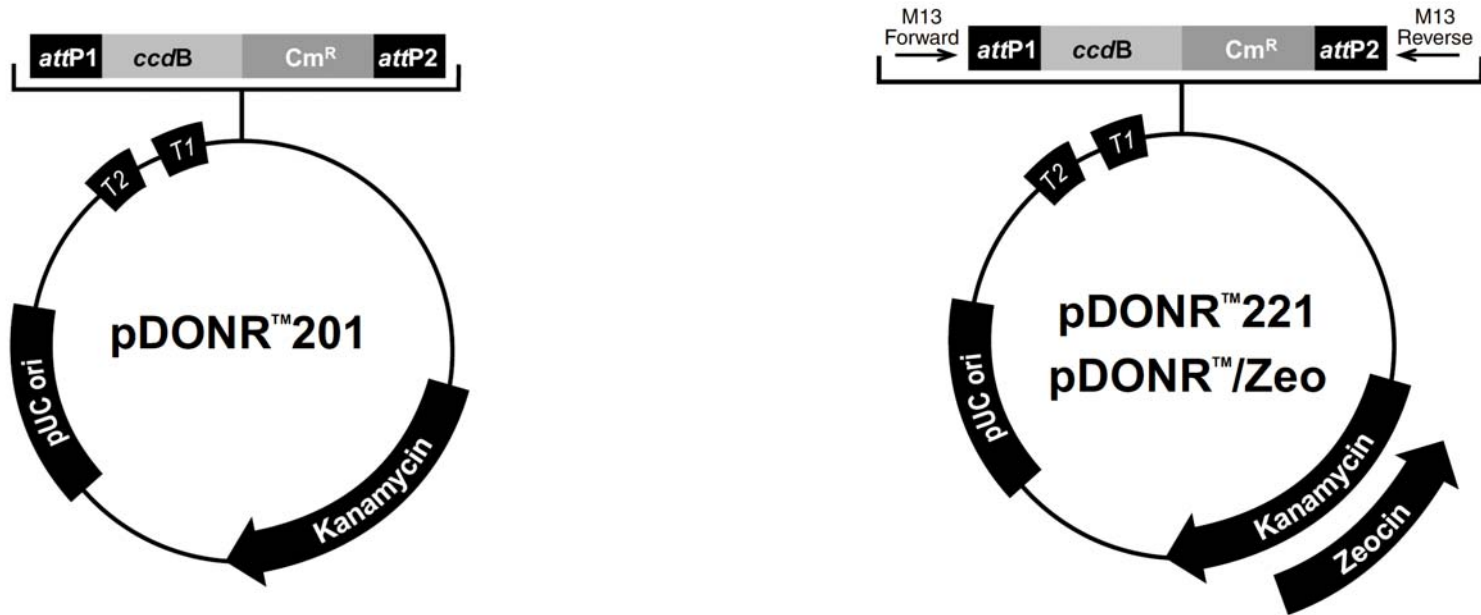
- Use when there are convenient sites to cut insert out of another plasmid
- Must cut out *ccdB* gene by using one of four RE sites flanking the *ccdB*
- Reading frame of insert must be considered, as well as downstream expression elements
- Various reading frames of pENTR vectors are available

pENTR™ 1A	Vector in reading frame 0
pENTR™ 2B	Vector in reading frame +1
pENTR™ 3C	Vector in reading frame +2
pENTR™ 4	Vector in reading frame 0; modified polylinker from 1A
pENTR™ 11	Contains both an <i>E. coli</i> (SD) and eukaryotic (Kozak) ribosome binding site

Options to Create Entry Clones III

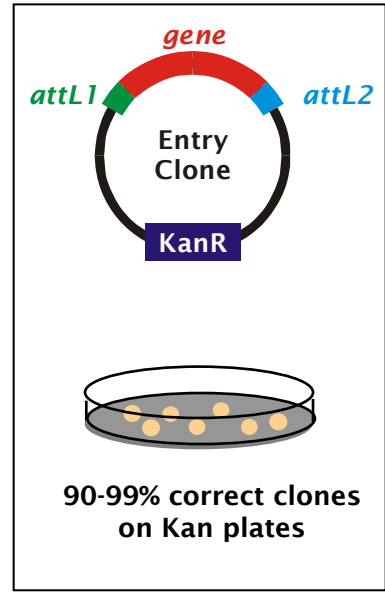
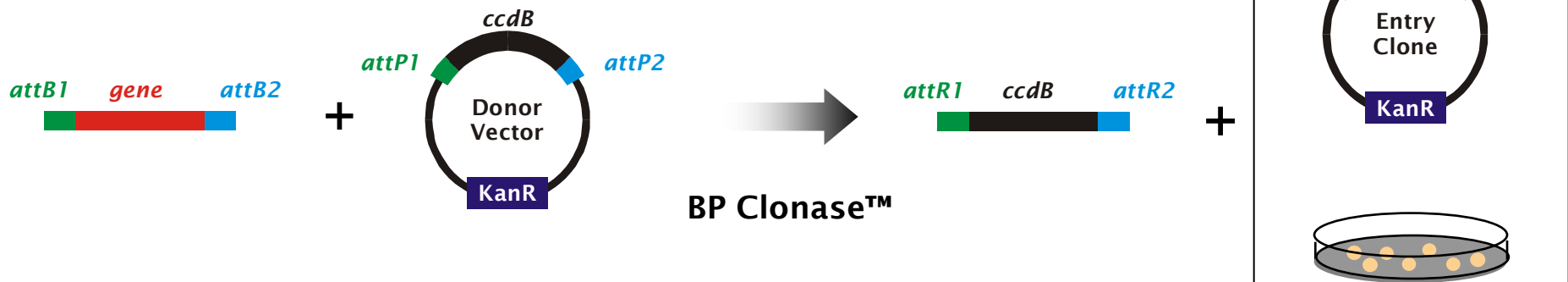


Map and Features of pDONR 201, pDONR221 and pDONR/Zeo



- *rrn* BT2 transcription termination sequence
- *rrn* BT1 transcription termination sequence
- *attP1* site
- *ccdB* gene
- chloramphenicol resistance gene
- *attP2* site
- kanamycin resistance gene } pDONR201 and pDONR221
- *pUC ori*
- T7 promoter / priming site } pDONR221 and pDONR/Zeo
- M13 reverse priming site } pDONR221 and pDONR/Zeo
- Zeocin resistance gene } pDONR/Zeo

BP Cloning – The Reaction





To be considered when designing PCR Primers:

- Sequences to facilitate Gateway cloning
- Sequences required for efficient expression of the native protein (i.e. Shine-Dalgarno or Kozak consensus)
- Whether or not you wish your PCR product to be fused in frame with an N- or C-terminal fusion tag

BP Cloning - Primer Design for PCR

- GGGG and the *attB1* sequence must be added to the 5'-primer (sense)
- GGGG and the *attB2* sequence must be added to the 3'-primer (antisense)

Forward Primer:

attB1

5' - GGGGACAAGTTTGTACAAAAAAGCAGGCTNNN...

Reverse Primer:

attB2

5' - GGGGACCACTTTGTACAAGAAAGCTGGGTNNN...

Designing *attB* Forward Primers



1) Forward Primer Design for [Native Expression](#)

5' -GGGG**ACAAGTTTGTACAAAAAGCAGGCT**^{Shine-Dalgarno}^{Kozak}**TTCGAAGGAGATAGAACCATGG**(18-25 gene-specific nucleotides)-3'

2) Forward Primer Design for [N-terminal Fusions](#)

5' -GGGG **ACA AGT TTG TAC AAA AAA GCA GGC** **TTC**(18-25 gene-specific nucleotides)-3'

Lys Lys

Designing *attB* Reverse Primers



1) Reverse Primer Design with no C-terminal fusion tag

5' - GGGG ACCACTTTGTACAAGAAAGCTGGGT CTA (18-25 gene-specific nucleotides) - 3'

attB2

2) Reverse Primer Design for C-terminal fusion tag

5' - GGGG - AC - CAC - TTT - GTA - CAA - GAA - AGC - TGG - GTN - (template-specific sequence) - 3'

attB2

Gateway® BP Clonase Reaction



<i>attB</i> -PCR Product	(30 - 300 ng)	1 – 10 μ l
Donor vector	(150 ng/ μ l)	2 μ l
5x BP Clonase Reaction Buffer		4 μ l
TE, pH 8.0		16 μ l

→ + 4 μ l BP Clonase enzyme mix

→ 25°C, over night

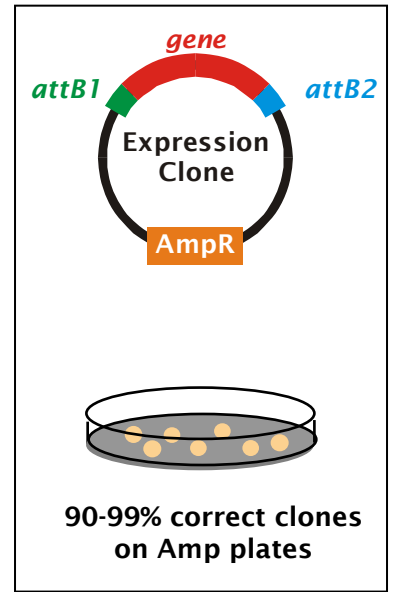
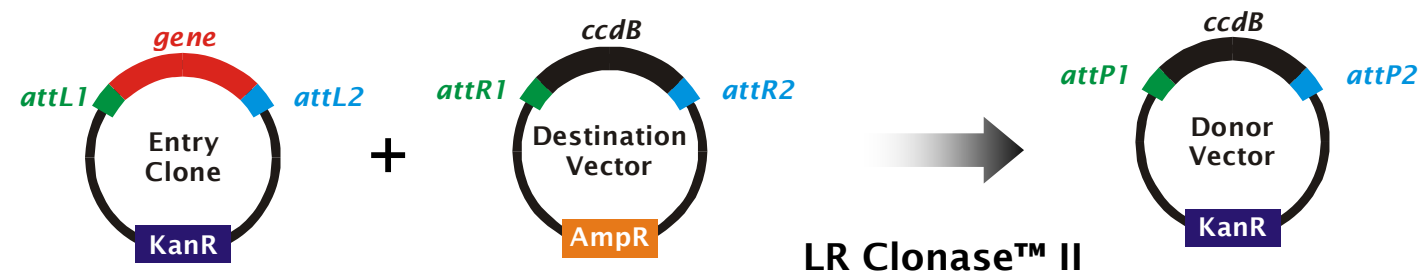
→ + 2 μ l Proteinase K

→ 37°C, 10 min

BP Clonase Mix: Reaction Buffer + Clonase Enzyme Mix

BP Clonase II Mix: Clonase Mix includes Reaction Buffer, (less expensive)

Gateway® LR-Recombination Reaction

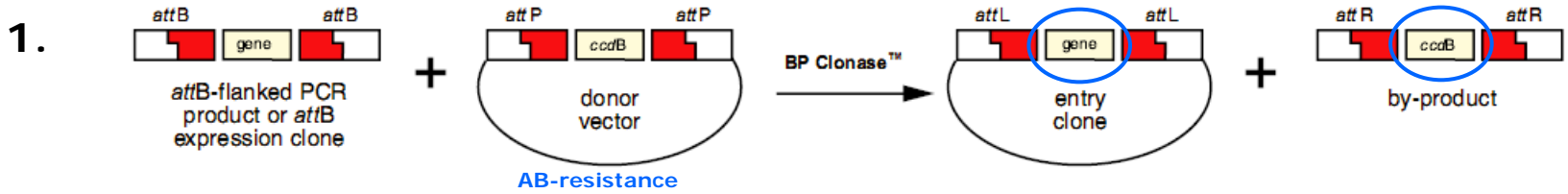


E. coli strains for transformation of Gateway constructs

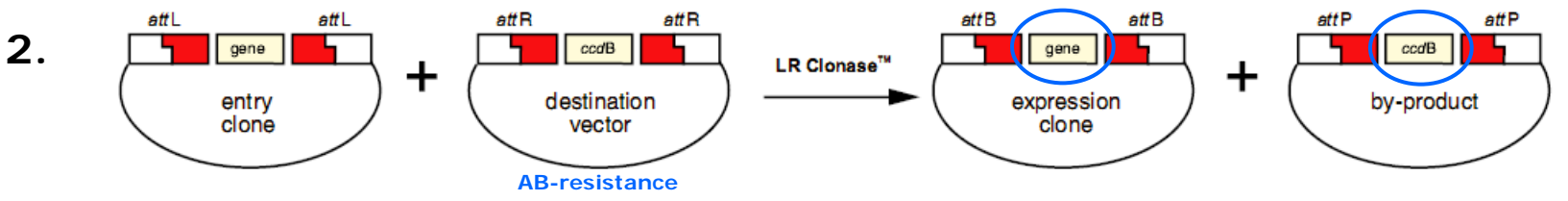


- | | |
|-----------------------------------|--|
| OmniMAX 2-T1[®] : | Selection of (positive) transformants,
„general“ cloning strain |
| DH5α: | Selection of (positive) transformants,
„general“ cloning strain |
| <i>ccdB</i>-survival : | To maintain Gateway – constructs without insert |
| DB3.1: | To maintain Gateway – constructs without insert |

Gateway[®]-Technology: Summary

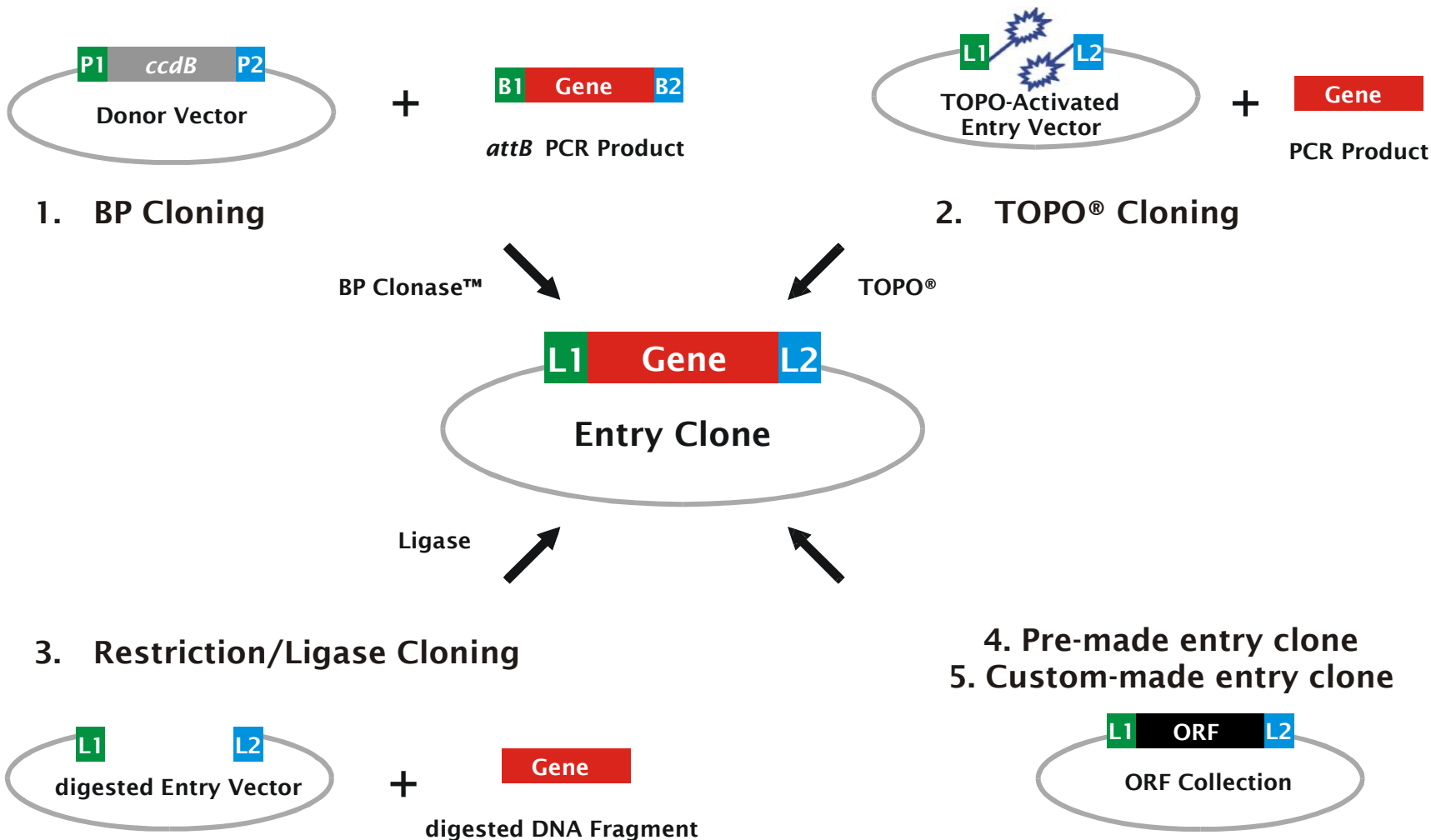


- **LR Reaction:** Facilitates recombination of an *attL* substrate (entry clone) with an *attR* substrate (destination vector) to create an *attB*-containing expression clone (see diagram below). This reaction is catalyzed by LR Clonase[™] enzyme mix.



	Pathway	Reaction	Catalyzed by...
1.	Lysogenic	$attB \times attP \rightarrow attL \times attR$	BP Clonase [™] (Int, IHF)
2.	Lytic	$attL \times attR \rightarrow attB \times attP$	LR Clonase [™] (Int, Xis, IHF)

Different ways to generate the entry clone





1. Overview and Entry Options
2. Destination Vectors

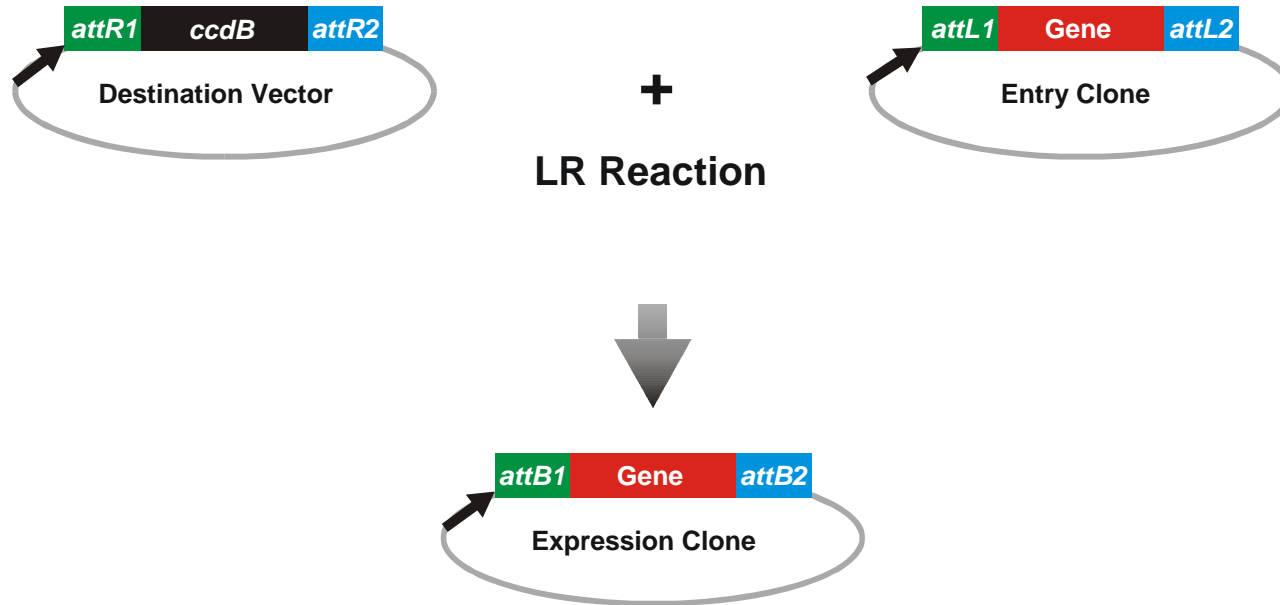


Fig. ©Invitrogen

destination vectors:

- have *attR* sites
- allow recombination with entry clones

Number of applications by Invitrogen



Table 2—Gateway® expression vector options.

Application	Gateway® Destination vector family
Protein array	Expressway™ Plus Expression system
Antibody or antigen production	Champion™ pET Expression systems
Protein expression in <i>E. coli</i>	pDEST™14, 15, 17, and 24 pET160 and pET161 DEST™ vectors
Protein expression in yeast	pYES2-DEST™52
Protein expression in insect cells	BaculoDirect™ C-term Expression Kit
Protein expression in mammalian cells (constitutive expression)	pcDNA® mammalian expression vector family
Protein expression in mammalian cells (regulated expression)	pT-REx-DEST30 and pT-REx-DEST31 vectors
Protein expression in mammalian cells (viral delivery)	ViraPower™ Lentiviral Expression Systems
Protein-protein interaction studies	ProQuest™ Two-Hybrid System using Gateway® technology
Localization	VividColors™ pcDNA GFP Destination vector family
RNAi	BLOCK-iT™ vector family
Reporter assay	GeneBLAzer™ pcDNA vector family

Destination vectors for protein expression

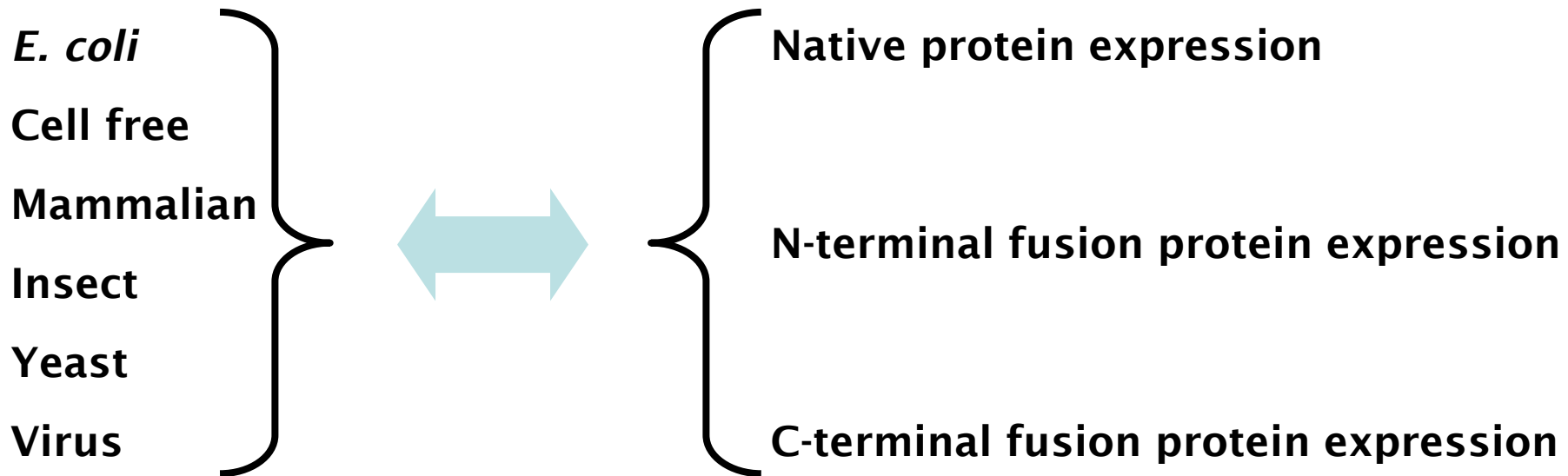
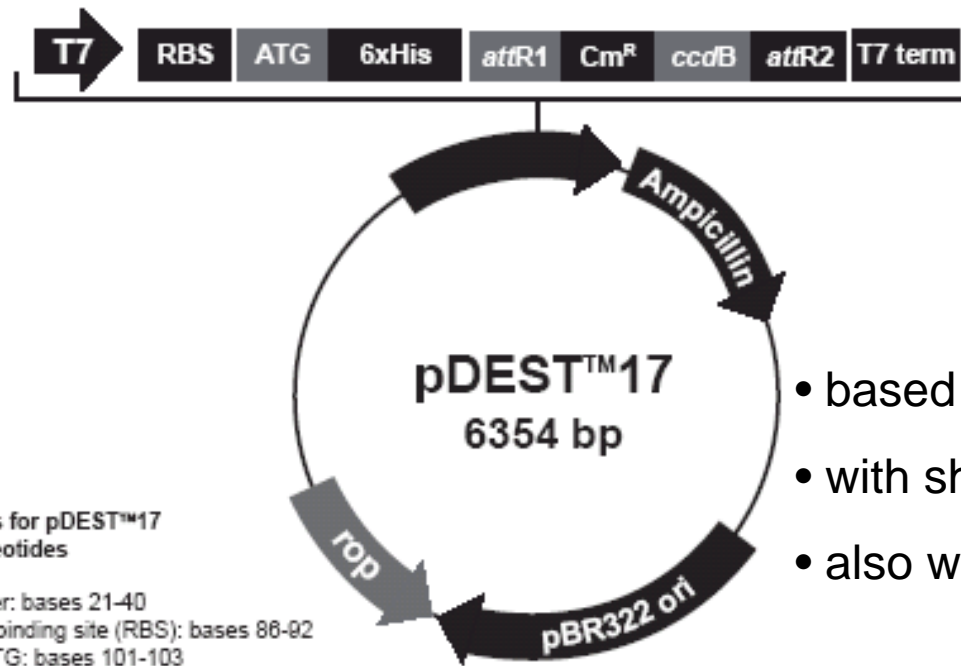


Fig. ©Invitrogen

E. coli: pDEST17



Comments for pDEST™17
6354 nucleotides

T7 promoter: bases 21-40

Ribosome binding site (RBS): bases 88-92

Initiation ATG: bases 101-103

6xHis tag: bases 113-130

attR1: bases 140-264

Chloramphenicol resistance gene (Cm^R): bases 373-1032

ccdB gene: bases 1374-1679

attR2: bases 1720-1844

T7 transcription termination region: bases 1855-1983

bla promoter: bases 2471-2569

Ampicillin (*bla*) resistance gene: bases 2570-3430

pBR322 origin: bases 3575-4248

ROP ORF: bases 4619-4810 (C)

C=complementary strand

- based on pET vectors
- with shine dalgarno sequence
- also with C-term Tag or GST-Tags

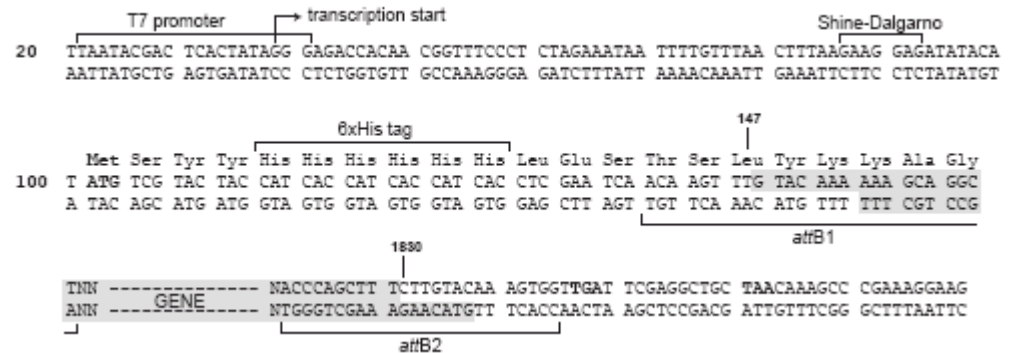
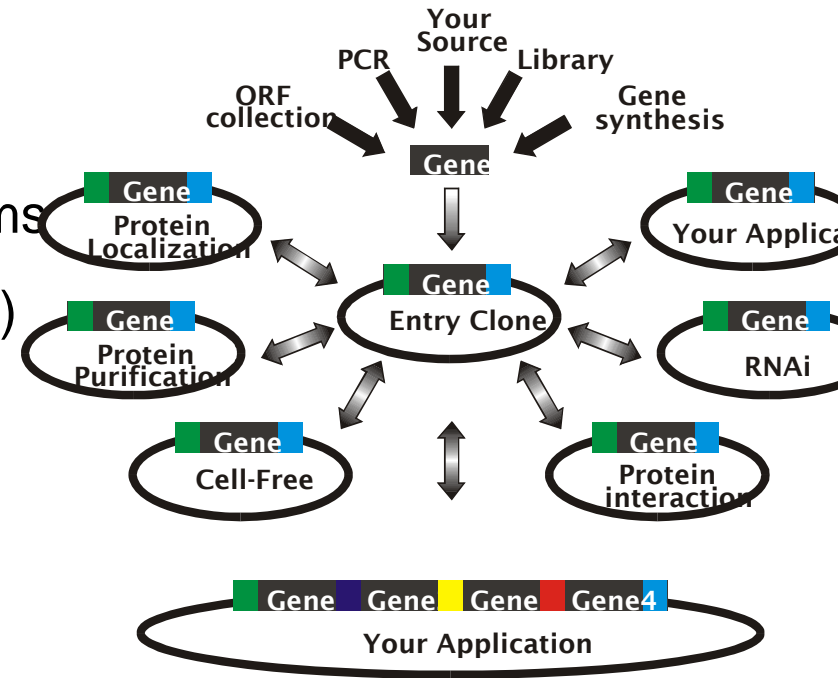


Figure & Sequence
©Invitrogen

overview about other destination vectors

- *in vitro* Transcription/ Translation
- protein expression in other organisms (e.g. yeast, mammalian, insect cells)
- Two-Hybrid System in Yeast
- RNAi



→ Creation of your own Gateway-vector:

conversion cassette

→ in plants: based on pCAMBIA

Fig. ©Invitrogen

Gateway® Conversion Kit

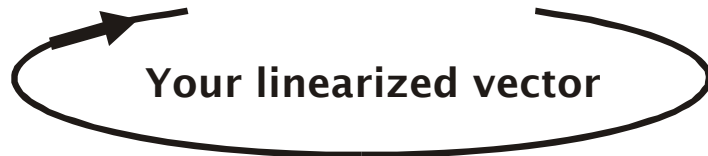


Gateway® Cassette

+



Promoter

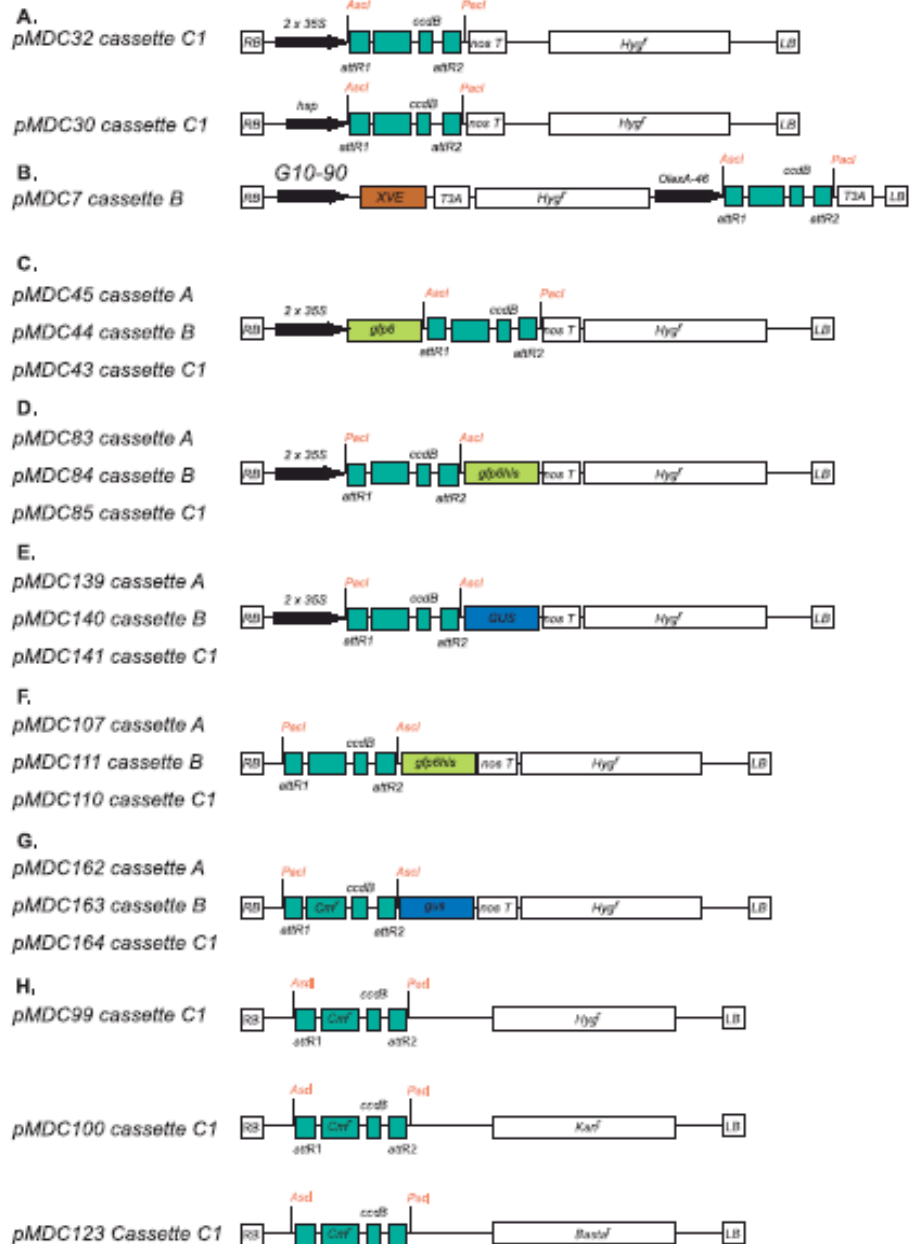
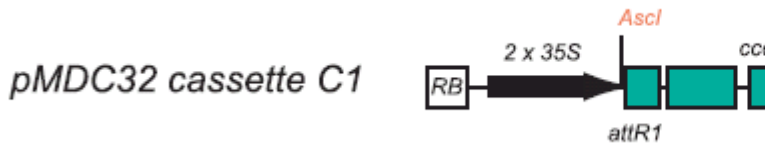


Conversion cassettes can be used with any vector or system, even the proprietary ones

examples for created plant transformation vectors



- pMDC Gateway Vector Series
(Curtis & Grossniklaus (2003) *Plant Phy*
→ Complementation



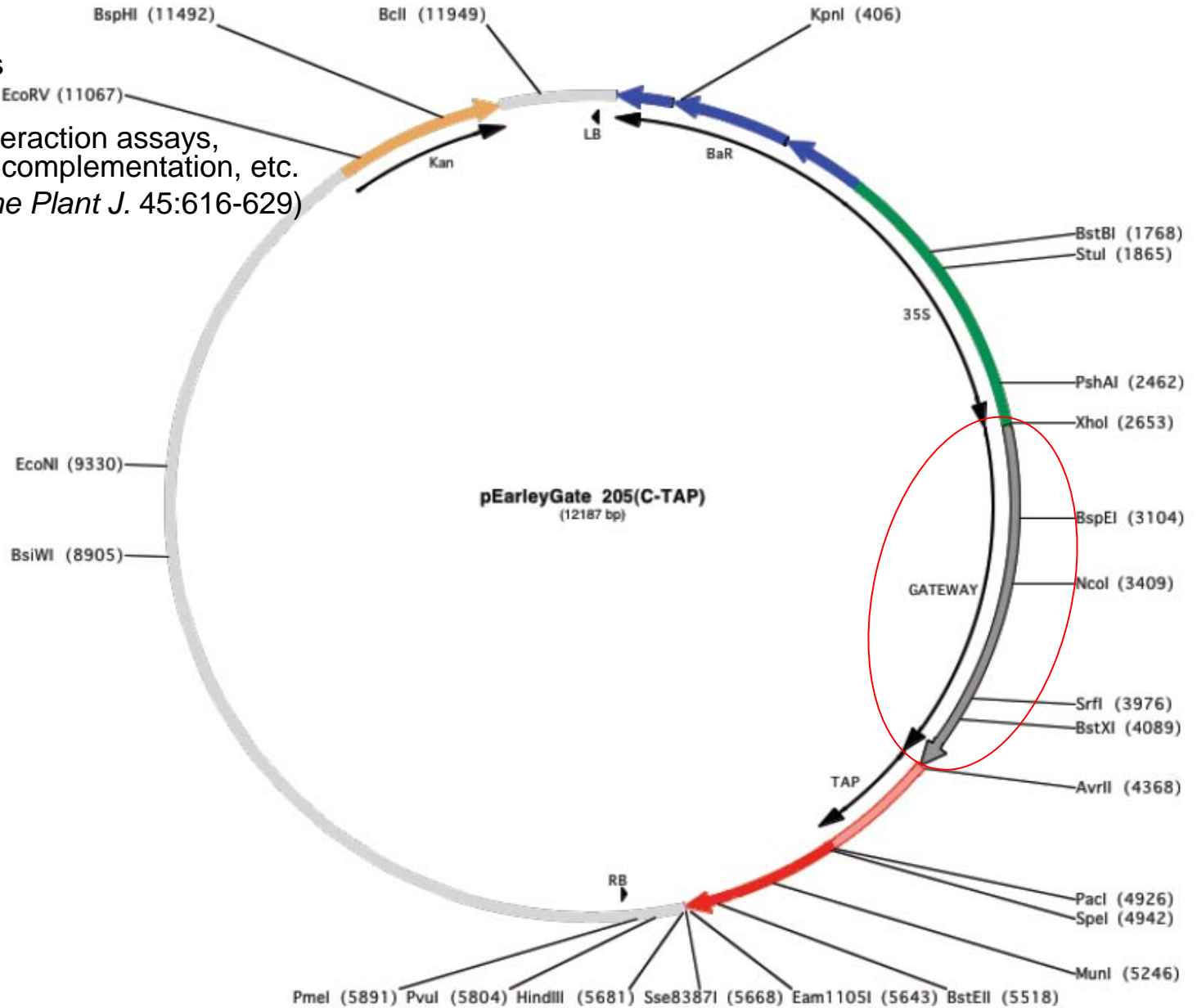
pEarleyGate Vector Series: pEarleyGate 205



pEarley Vector Series
by Pikaard laboratory

→ Purification and Interaction assays,
Fluorescent Imaging, complementation, etc.

(Earley *et al.* (2006) *The Plant J.* 45:616-629)



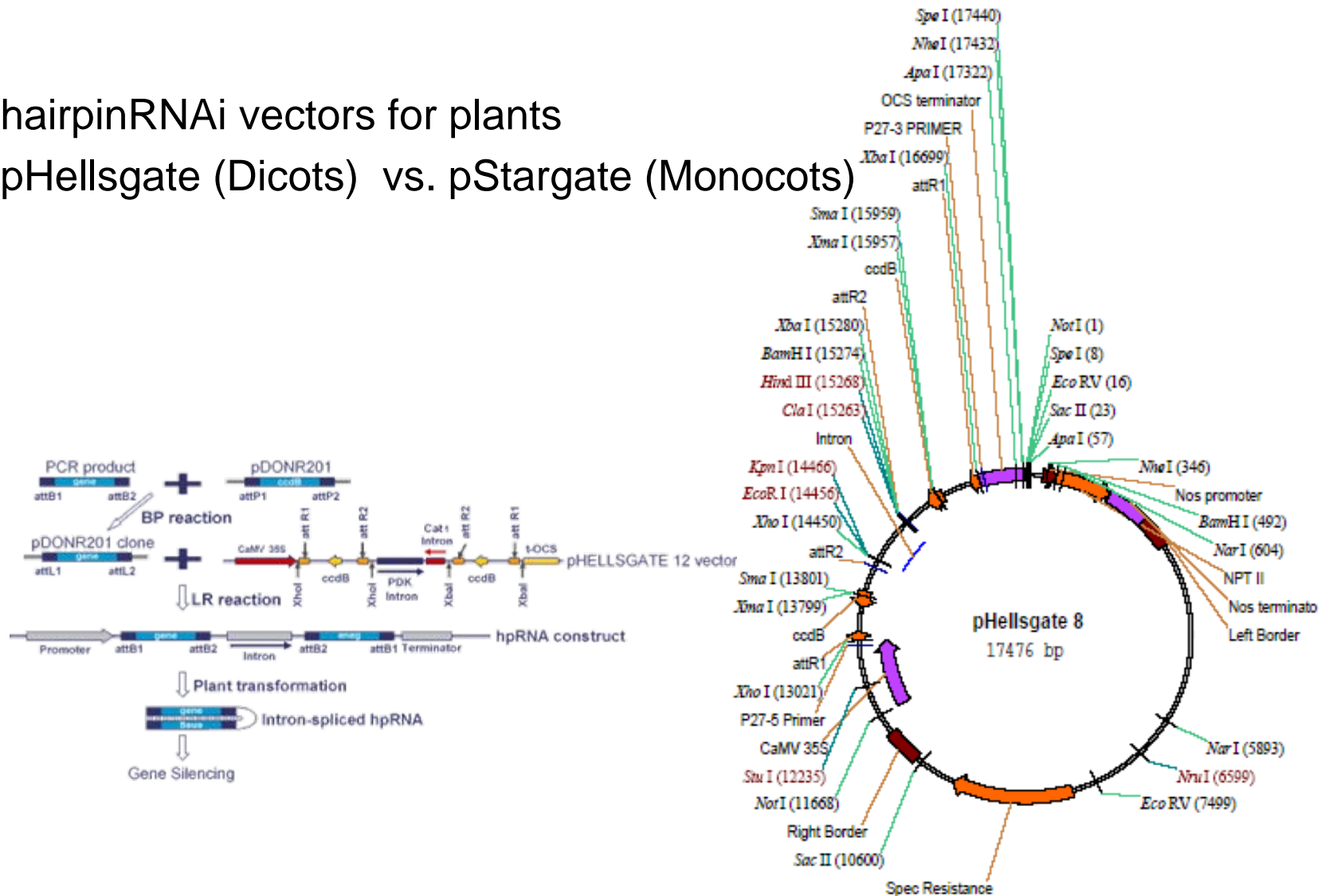
pEarleyGate Vector Series: overview



Plasmid name	Description
pEarleyGate 100	35S-Gateway-OCS 3'
pEarleyGate 101	35S-Gateway-YFP-HA tag-OCS 3'
pEarleyGate 102	35S-Gateway-CFP-HA tag-OCS 3'
pEarleyGate 103	35S-Gateway-GFP-His tag- OCS 3'
pEarleyGate 104	35S-YFP-Gateway-OCS 3'
pEarleyGate 201	35S-HA tag-Gateway-OCS 3'
pEarleyGate 202	35S-FLAG tag-Gateway-OCS 3'
pEarleyGate 203	35S-Myc tag-Gateway-OCS 3'
pEarleyGate 204	35S-AcV5 tag-Gateway-OCS 3'
pEarleyGate 205	35S-Gateway-TAP tag-OCS 3'
pEarleyGate 301	no promoter-Gateway-HA tag-OCS 3'
pEarleyGate 302	no promoter-Gateway-FLAG tag-OCS 3'
pEarleyGate 303	no promoter-Gateway-Myc tag-OCS 3'
pEarleyGate 304	no promoter-Gateway-AcV5 tag-OCS 3'

RNAi in planta: pHellsgate series

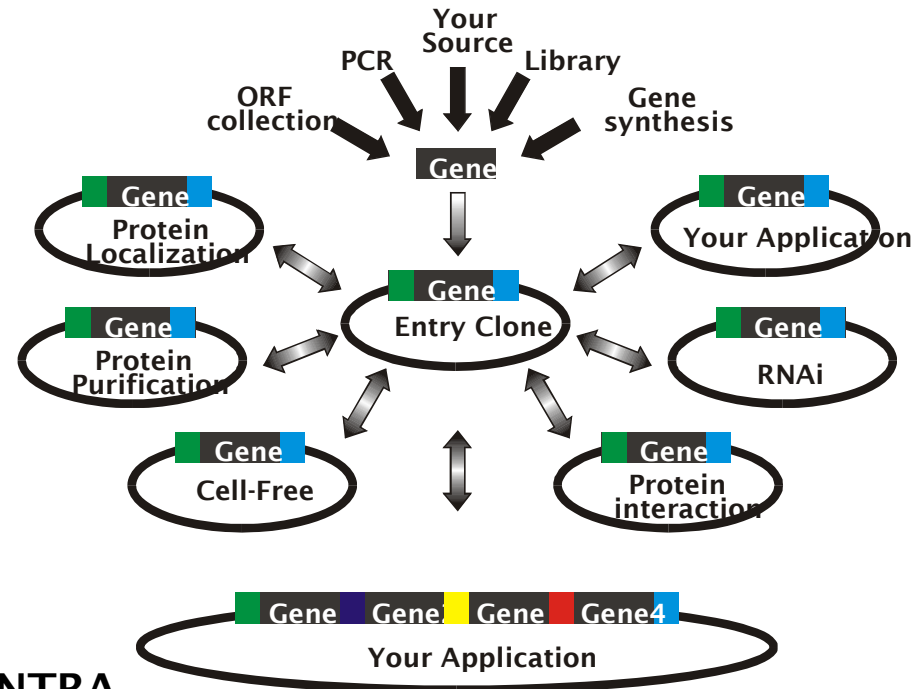
- hairpinRNAi vectors for plants
- pHellsgate (Dicots) vs. pStargate (Monocots)



conclusion of The Gateway[®] Cloning System

PRO

- Directional cloning
- Maintains reading frame
- No restriction enzymes
- No ligation
- 1 hour, room-temperature reaction with >99% efficiency
- No re-sequencing
- Compatible with automation
- Reversible reactions



CONTRA

- Primer
- dependance on Invitrogen (LR/BP reaction mix)
- antibiotics resistance

Thank you for
your attention!



Weiterführende Internet-Seiten der Firma Invitrogen:

<http://www.invitrogen.com/site/us/en/home/Products-and-Services/Applications/Cloning/Gateway-Cloning.html>

<http://www.invitrogen.com/etc/medialib/en/filelibrary/pdf/Brochures.Par.38170.File.dat/B-074573-Gateway.pdf>

<http://www.invitrogen.com/site/us/en/home/Products-and-Services/Applications/Cloning/Gateway-Cloning/GatewayC-Misc/Online-Seminars.html>