Phylogenetic relationships in *Carex*, subgenus *Vignea* (Cyperaceae), based on ITS sequences

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Abstract. To evaluate the sectional classification in Carex, subgenus Vignea, the ITS region of 58 species of 20 sections was analyzed with Neighbor Joining (NJ) and Markov chain Monte Carlo (MCMC) methods. Sections Dioicae, Physodeae and Ovales are found to be monophyletic, with C. bohemica well integrated in the section Ovales. Section Heleonastes turns out to be monophyletic, if C. canescens is treated separately in section Canescentes. Section Elongatae is monophyletic, but C. remota is placed in section Remotae and C. bromoides in section Deweyanae. In both analyses, six representatives of section Arenariae cluster together in a terminal group, whereas C. disticha, C. repens and C. siccata form a basal cluster. C. maritima, as the only member of section Incurvae, shares this basal position. C. chordorrhiza is ascribed to section Chordorrhizeae and not ascribed to the paraphyletic section Divisae. C. vulpina and C. otrubae are assigned to section Vulpinae and separated from the heterogeneous section Stenorhynchae. The other members of sections Divisae, Muehlenbergianae, Multiflorae and Stenorhynchae are scattered throughout the trees. The representatives of section Foetidae are dispersed in both analyses, section Paniculatae appears to be non-monophyletic in the molecular results as well.

The subgenus appears subdivided in at least four larger subgroups in all analyses. Whereas these

subgroups are strongly supported, the relationships between these subgroups remain only poorly resolved.

Key words: Bayesian analysis, *Carex*, *Vignea*, ITS, molecular phylogeny, systematics.

Carex subgenus Vignea (Beauv.) Nees is morphologically characterized by the lack of a prophyll surrounding the base of the axis of partial inflorescence (cladoprophyll), mostly bisexual spikes and usually 2, rarely 3 stigmas. It has been regarded as a natural lineage within the genus Carex L. since the early systematic studies in the 19th century (e.g. Schweinitz 1825, Dumortier 1827, Fries 1835, Kunth 1837, Tuckerman 1843, Drejer 1844). Nearly a quarter of the species of *Carex* are members of subgenus Vignea. In North America it comprises about one third of the recognized Carex species, in Eurasia estimations reach from 25 to 30% (Ball 1990, Egorova 1966). In the worldwide monograph of Georg Kükenthal (1909) subgenus Vignea contains 20 sections with 140 species in his broad species concept. Since then, number of species has at least tripled, due to a narrower

species concept and to new findings especially outside Europe. For North America, 168 species in 17 sections are described (Ball and Reznicek 2002), the Flora of Russia and adjacent countries (Egorova 1999) covers 320 species in 25 sections.

For subgenus Carex, the potential of the ITS region for phylogenetic interpretations on the sectional level was recently demonstrated (Roalson et al. 2001, Hendrichs et al. 2004). For subgenus Vignea, the available molecular data are scarce. Recent molecular studies focused mainly on family and subfamily relationships (Muasya et al. 1998, Yen and Olmstead 2000) or on species of subgenus Carex (Waterway and Olmstead 1998, Starr et al. 1999, Roalson et al. 2001). For better understanding of sectional circumscriptions in subgenus Vignea we analyzed 58 species mainly from northern Europe including sequences of 12 species derived from GenBank (see Table 1).

Materials and methods

Plant collection and DNA extraction. The analyzed species of *Carex* are listed in Table 1. Assignment of sections and subsections corresponds mainly to the concept of Kükenthal (1909). Total genomic DNA was isolated from fresh or dried leaf tissue either by crushing the plant material in liquid nitrogen with a micro pestle or by shaking the samples for 3 min at 30 Hz (Mixer Mill MM 300, Retsch, Haan, Germany). DNeasy Plant Mini Kit (Qiagen, Hilden, Germany) was used following the manufacturer's protocol.

PCR and sequencing. The ITS region (ITS1, 5.8 S, ITS2; about 700 bp) was amplified with the primer pair ITSL (Hsiao et al. 1995), ITS5 and ITS4 (White et al. 1990) or ITS5i and ITS4i (Roalson et al. 2001). PCR was performed as described in Hendrichs et al. (2004). The PCR product was purified with QIAquick PCR Purification Kit (Qiagen, Hilden, Germany). The dsDNA obtained was sequenced directly on both strands using the ABI PRISM Big DyeTM Terminator Cycle Sequencing Ready Reaction Kit (PE Applied Biosystems) on an automated sequencer (ABI 373A, PE Applied Biosystems and LICOR 4000). The

sequences of both strands were combined and proof-read with SequencherTM 4.1 software (Gene Codes Corp., Michigan) and BioEdit (Hall 1999). The sequences reported in this study have been deposited in GenBank (see Table 1). The alignment contained 614 nucleotide sites. After removing ambiguously aligned positions (220–233), 600 sites remained for analyses with 234 variable sites (ITS1: 129, 5.8S: 8, ITS2: 97). The ingroup alone contained 220 variable sites. The alignment is available upon request.

Phylogenetic analysis. DNA sequences were aligned using Clustal X (Jeanmougin et al. 1998). Some manual corrections were done in Se-Al v2.0a7b (Rambaut 2001). The likelihood ratio test as implemented in Modeltest 3.0 (Posada and Crandall 1998) selected TrN + G (Swofford et al. 1996) as substitution model (details below). A Bayesian method of phylogenetic inference using a Metropolis-coupled Markov chain Monte Carlo (MCMC) approach was carried out as implemented in the computer program MrBayes (Huelsenbeck and Ronquist 2001) with GTR + I + G(Swofford et al. 1996) as substitution model. Four incrementally heated simultaneous Monte Carlo Markov chains were run over 2 000 000 generations. Trees were sampled every 100th generation, resulting in an overall sampling of 20 000 trees. To obtain estimates for the a posteriori probabilities, a 50% majority rule consensus tree was computed from those trees that were sampled after the process had reached stationarity (burnin = 2000). This Bayesian approach of phylogenetic analysis was repeated five times, always using random starting trees and random starting values for the model parameters to test the reproducibility of the results. Branch lengths were estimated under the maximum likelihood criterion using the same substitution model in PAUP 4.0b10 (Swofford 2002).

Neighbor joining analysis (Saitou and Nei 1987) was done with PAUP 4.0b10 (Swofford 2002) using genetic distances estimated under the maximum likelihood criterion and TrN + G as substitution model with the following settings: base frequencies A = 0.235075, C = 0.273445, G = 0.281677, T = 0.209804;rate matrix AC = 1.00000, AG = 2.25212, AT = 1.00000, CG = 1.00000, CT = 6.34211, GT = 1.00000;gamma distribution shape parameter = 0.357885. Support for internal nodes was estimated by 1000 neighbor joining bootstrap replicates under the

M. Hendrichs et al.: Carex, subgenus Vignea (Cyperaceae)

Species	$Locality/Voucher^{\dagger}$	GenBank accession no.
Carex alma L. H. Bailey	USA, California*	AF285025
Carex appropinquata Schum.	Germany; HMH 3358	AY280549
Carex arenaria L.	Germany; SMC 126	AY280529
Carex athrostachya Olney	USA, Washington; JM 132	AY280539
Carex bicknellii Britton	USA, Michigan*	AF285039
Carex bohemica Schreb.	Germany; SMC 75	AY280532
Carex bonplandii Kunth	Costa Rica; HeRB 6108	AY280563
Carex brizoides L.	Germany; HMH 503	AY280546
Carex bromoides Schkuhr	USA, Tennessee; JE	AY280534
Carex brunnescens (Pers.) Poir.	Sweden; HMH 2776	AY280567
Carex canariensis Kük.	Spain; HMH 2938	AY280558
Carex canescens L. ^a	USSR, Siberia*	AF284990
Carex canescens L. ^b	France; HMH 2282	AY280550
Carex chordorrhiza Ehrh.	Sweden; HMH 2804	AY280568
Carex curvata Knaf	Germany; SMC 131	AY280530
Carex davalliana Smith	Germany; HMH 1413	AY280542
Carex densa L. H. Bailey	USA, Washington; JM 161	AY280538
Carex diandra Schrank	Denmark; HMH 1931	AY280551
Carex dioica L.	Germany; HeRB 2185	AY280543
Carex disticha Lam.	Germany; FO 8396	AY280571
Carex divisa Huds.	France; FO 18408	AY280552
Carex divulsa Stokes	Germany; HMH 1792	AY280553
Carex duriuscula C.A. Mey.	Canada,**	AF027436
Carex echinata Murray	Denmark; HMH 1920	AY280559
Carex elongata L.	Germany; FO 9351	AY280560
Carex foetida All.	France; HMH 2292	AY280544
Carex fracta Mack.	USA, California*	AF285030
Carex heleonastes Ehrh.	Germany; HMH 2946	AY280566
Carex hoodii Boott	USA, Washington; JM 129	AY280537
Carex jonesii L. H. Bailey	USA, California*	AF285038
Carex lachenalii Schkuhr	Sweden; HMH 2692	AY280564
Carex leavenworthii Dewey	USA, Texas*	AF285033
Carex leersii F.W. Schultz	France; HeRB 3031	AY280554
Carex ligerica Gay	Germany; SMC 120	AY280531
Carex loliacea L.	Sweden; HMH 2955	AY280565
Carex macloviana d'Urv.	Sweden; HMH 2957	AY280562
Carex macrocephala Willd.	USA, Oregon*	AF285017
Carex macrorrhiza Boeck.	Argentina, Santa Cruz*	AF285018
Carex maritima Gunn.	Switzerland; JM 146	AY280570
Carex muricata L.	Estonia, Muhu Island*	AF285036
Carex muskingumensis Schw.	Canada; SMC 181	AY280541
Carex otrubae Podp.	Germany; HMH 1776	AY280556
Carex ovalis Good.	Germany; HMH 1770	AY280561
Carex paniculata L.	Switzerland; HMH 2865	AY280557
<i>Carex parallela</i> (Laest.) Sommerf.	Sweden; HMH 2780	AY280545
Carex pinetorum Liebm.	Bolivia; LPB	AY280540

 Table 1. Species analyzed in this study

Species	$Locality/Voucher^{\dagger}$	GenBank accession no.
Carex praecox Schreb.	Germany; SMC 149	AY280527
Carex pseudobrizoides Clavaud	Germany; SMC 97	AY280526
Carex remota L.	Germany; HMH 3354	AY280548
Carex repens Bell.	Poland; SMC 93	AY280528
Carex siccata Dewey	USA, Washington; JM 125	AY280536
Carex spicata Huds.	Germany; HeRB 624	AY280555
Carex stenophylla Wahlenb.	Spain; JM 9772	AY280535
Carex stipata Muehlenb.	USA, Wisconsin; JE	AY280533
Carex vallicola Dewey	USA, Wyoming; FO 31030	AY280569
Carex vernacula L. H. Bailey	USA, Oregon*	AF285022
Carex vulpina L.	Germany; HMH 3359	AY280547
Carex vulpinoidea Michx.	USA, Texas*	AF284968
Kobresia capillifolia (Decne.) C. B. Clarke	China, Xinjiang*	AF284984
Kobresia myosuroides (Vill.) Fiori & Paol.	USSR, Siberia*	AF284985
Kobresia sibirica (Turcz. ex Ledeb.) Boeck.	USSR, Siberia*	AF284986

 Table 1 (continued)

[†] Acronyms of herbaria and collections: GOET: Herbarium University Göttingen; HeRB: R. Berndt (private collection); HMH: M. Hendrichs (private collection); FO: F. Oberwinkler (private collection); JE: Herbarium Haussknecht, Jena; JM: J. Müller (private collection); LPB: Herbarium Nacional de Bolivia; SMC: S. Michalski (private collection); TUB: Herbarium Tubingense; WM: W. Maier (private collection); * Origin of sequence: Roalson et al. 2001

** Origin of sequence: Starr et al. 1999

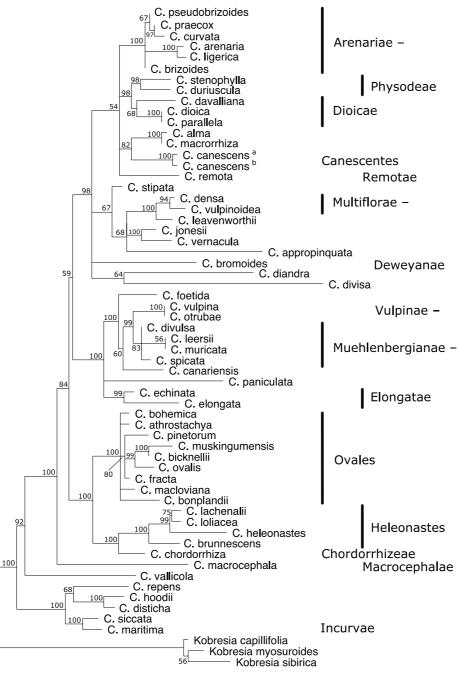
same model settings. The unrooted phylograms from neighbor joining and MCMC analyses were rooted with three species of the genus *Kobresia* Willd. from GenBank.

Results

The different runs of Bayesian phylogenetic analysis yielded consistent results. Stationarity of the Markov chains was reached after approximately 200 000 generations of trees, i.e. after 2000 trees had been sampled. Thus, we discarded the first 2000 trees and included 18 000 sampled trees in the 50% majority rule consensus tree of each run. One of them is given in Fig. 1. The phylogram obtained by the NJ analysis is shown in Fig. 2. In both analyses the supported clusters contain the same species, and the tree topology of the MCMC analysis correlates with that of the NJ analysis. In general, statistical support is higher in the MCMC topology than in the NJ topology (compare Fig. 1 with Fig. 2). Parsimony analyses revealed consistent results, but with lower statistical significance (Hendrichs et al. 2003a).

Rooted with three species of the genus *Kobresia*, the members of subgenus *Vignea* group as a highly supported monophyletic lineage. The use of *Kobresia* as outgroup for our analyses is justified through the results of molecular investigations in the relationship of the genus *Carex* (comp. Yen and Olmstead 2000, Roalson et al. 2001, Hendrichs et al. 2003b).

The sectional delimitations within subgenus *Vignea* have been of great difficulty and constant rearrangement since the early approaches. Of the 20 traditionally accepted sections represented in our analyses only sections *Dioicae*, *Physodeae* and *Ovales* are found to be monophyletic. *C. bohemica*, often placed in section *Schellhammeria*, appears fully integrated in section *Ovales*. Section



- 0.005 substitutions/site

Fig. 1. Bayesian inference of phylogenetic relationships within *Carex* subgenus *Vignea*. Metropolis-coupled Markov chain Monte Carlo analysis of an alignment of nuclear sequences from the ITS region using the general time reversible model of DNA substitution with gamma distributed substitution rates and estimation of variant sites. 50% majority rule consensus tree from 18 000 trees that were sampled after the process had reached stationarity. The topology was rooted with three species of the genus *Kobresia*. The numbers on branches are estimates of *a posteriori* probabilities. Branch lengths were estimated using Maximum Likelihood and are scaled in terms of expected numbers of nucleotide substitutions per site. The groups marked with "—" do not include all analyzed members of the specified section

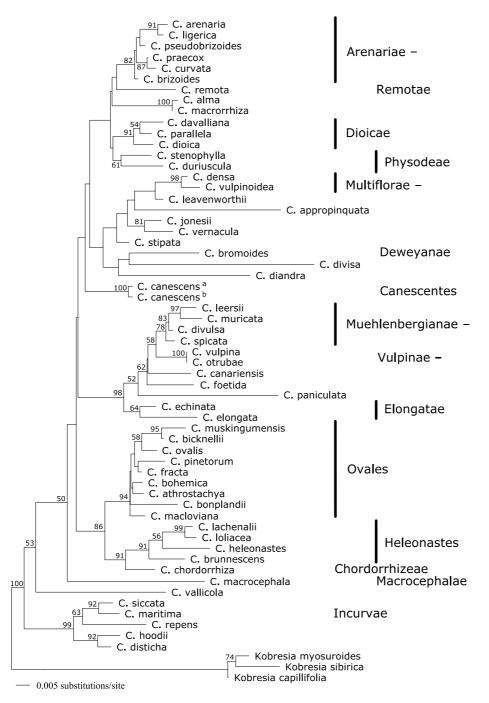


Fig. 2. ITS phylogram of *Carex* subgenus *Vignea* obtained by neighbor joining analysis using TrN + G as substitution model (parameters are given in the text). The topology was rooted with three species of the genus *Kobresia*. Percentage bootstrap values of 1000 replicates are given at each furcation values smaller than 50% are not shown. Branch lengths are scaled in terms of expected numbers of nucleotide substitutions per site. Groups marked with "–" do not include all analyzed members of the specified section

Macrocephalae is represented in our analyses only by *C. macrocephala*. Section *Heleonastes* is found to be monophyletic, if *C. canescens* is treated in separated section *Canescentes*. Section *Elongatae* is marked monophyletic, whereby *C. remota* is placed in section *Remotae* and *C. bromoides* is referred to section Deweyanae. These two sections are represented by one species each in our dendrograms. In both analyses, six representatives of section Arenaria constitute a well supported group. C. disticha, C. repens and C. siccata, traditionally treated in section Arenariae, fall into a basal cluster. C. maritima, as the only member of section Incurvae, shares this basal position in our dendrograms. C. chordorrhiza is referred to section Chordorrhizeae and is not treated in the paraphyletic section Divisae. C. vulpina and C. otrubae are assigned to section Vulpinae and separated from the heterogeneous section Stenorhynchae. Circumscription of sections Divisae, Muehlenbergianae, Multiflorae and Stenorhynchae was always regarded as very difficult and artificial in larger parts (e.g. Kükenthal 1909, Ball and Reznicek 2002; comp. Table 2). The members of these sections are mainly scattered throughout the trees. If two or three species cluster together, these are labeled in our dendrograms as core-groups, marked with a "-" to indicate that not all analyzed members of the specified section are included. The representatives of section *Foetidae* are dispersed in both analyses. According to our analyses, section Paniculatae appears to be non-monophyletic as well, even though well-defined by gross morphology.

The ITS region is useful in defining sections within subgenus Vignea and reveals at least four larger subgroups comprising different sections. In a first subgroup species traditionally ascribed to section Arenariae cluster together with C. hoodii and C. maritima. A second well supported cluster includes members of sections Ovales and Heleonastes together with C. chordorrhiza. Species of sections Muehlenbergianae, *Vulpinae* and *Elongatae* form a third well supported subgroup with C. canariensis, C. foetida and C. paniculata included. The largest subgroup comprises species of sections Arenariae, Physodae, Dioicae and Multiflorae together with C. remota, C. alma, C. macrorrhiza, C. leavenworthii, C. appropinquata, C. jonesii, C. vernacula, C. stipata, C. bromoides, C. divisa, *C. diandra* and *C. canescens*. However, the relationships between these subgroups are only weakly supported.

Chromosome numbers of the species studied are listed in Table 2, giving the chromosome counts available in literature.

Discussion

The sections and species discussed are arranged in order of position in Fig. 1, starting at the base of the dendrogram.

Sections Incurvae, Macrocephalae and related species. A highly supported group in both analyses (a posteriori probability 100%, bootstrap value 99%) comprises C. hoodii, C. maritima and three species ascribed to section Arenariae by Kükenthal (1909): C. siccata, C. repens and C. disticha. Carex siccata forms a monophyletic group with C. maritima which was placed in the monotypic section Incurvae by Kükenthal (1909). C. disticha is originally an Eurasian endemic, but introduced to North America. It shows closer affinity to C. hoodii, which Kükenthal (1909) ascribed to the large and inhomogeneous section Muehlenbergianae. These two subgroups are highly supported (a posteriori probability 100%, bootstrap value 92%). Although different sectional classifications for C. maritima and C. disticha have been proposed (e.g. Egorova 1966, Hylander 1966, Chater 1980, Ball and Reznicek 2002), the group as a whole is not congruent with any existing sectional concept. C. repens is a local endemic species with three disjunct areals in Central Europe, a hybrid origin was sometimes suggested (e.g. Chater 1980). It differs morphologically from C. arenaria only by a greater number of spikes and the absence of a scalelike prophyll at the base of spikes. In molecular dendrograms, C. arenaria and other members of section Arenariae cluster together as a group at the greatest possible distance in a terminal branch. Interestingly, this group as a whole is characterized by a specific indel in alignment-position 10 and a characteristic sequence pattern in positions 120–123.

Species	Section	Chromos. no. $(2n)^*$
Carex alma L. H. Bailey	Multiflorae Kunth	
Carex appropinquata Schum.	Paniculatae Kunth	64
Carex arenaria L.	Arenariae Kunth	56, 58, 64
Carex athrostachya Olney	Ovales Kunth	68
Carex bicknellii Britton	Ovales Kunth	76, 78
Carex bohemica Schreb.	Schellhammeria (Moench) Kunth	62, 64, 80
Carex bonplandii Kunth	Ovales Kunth	
Carex brizoides L.	Arenariae Kunth	58
Carex bromoides Schkuhr	Deweyanae Tuck.	64, 66, 68
Carex brunnescens (Pers.) Poir.	Heleonastes Kunth	56
Carex canariensis Kük.	Muehlenbergianae Tuck.	58
Carex canescens L. ^a	Canescentes Fries	54, 56
Carex chordorrhiza Ehrh.	Chordorrhizeae Fries	62
Carex curvata Knaf	Arenariae Kunth	58
Carex davalliana Smith	Dioicae Tuck.	46
Carex densa L. H. Bailey	Multiflorae Kunth	
Carex diandra Schrank	Paniculatae Kunth	50, 54, 60
Carex dioica L.	Dioicae Tuck.	52
Carex disticha Lam.	Arenariae Kunth	62
Carex divisa Huds.	Divisae Christ	58, 60, 62
Carex divulsa Stokes	Muehlenbergianae Tuck.	56, 58
Carex duriuscula C.A. Mey.	<i>Physodeae</i> Christ ex Kük.	52
Carex echinata Murray	<i>Elongatae</i> Kunth	56, 58
Carex elongata L.	<i>Elongatae</i> Kunth	56
Carex foetida All.	<i>Foetidae</i> Tuck.	58
Carex fracta Mack.	Ovales Kunth	
Carex heleonastes Ehrh.	Heleonastes Kunth	56
Carex hoodii Boott	Muehlenbergianae Tuck.	58, 60
Carex jonesii L. H. Bailey	Stenorhynchae Holm	
Carex lachenalii Schkuhr	Heleonastes Kunth	58, 62, 64, 74
Carex leavenworthii Dewey	Muehlenbergianae Tuck.	56, 62, 61, 71
Carex leersii F.W. Schultz	Muehlenbergianae Tuck.	58
Carex ligerica Gay	Arenariae Kunth	58
Carex loliacea L.	Heleonastes Kunth	54
Carex macloviana d'Urv.	Ovales Kunth	82, 86
Carex macrocephala Willd.	Macrocephalae Kük.	74
Carex macrorrhiza Boeck.	Divisae Christ	, .
Carex maritima Gunn.	Incurvae Kük.	60
Carex muricata L.	Muehlenbergianae Tuck.	56, 58
Carex muskingumensis Schw.	Ovales Kunth	80
Carex otrubae Podp.	Vulpinae (Carey) Christ	58, 60
Carex ovalis Good.	Ovales Kunth	62, 64, 66, 68
Carex paniculata L.	Paniculatae Kunth	60, 62, 64
<i>Carex parallela</i> (Laest.) Sommerf.	Dioicae Tuck.	43, 44
Carex pinetorum Liebm.	Ovales Kunth	тэ, тт
Carex praecox Schreb.	Arenariae Kunth	58
Carex pseudobrizoides Clavaud	Arenariae Kunth	50
Curex pseudobrizoides Clavaud	Arenariae Kultul	62

Remotae Aschers.

62

Table 2. Species, sections, and chromosome numbers. The classification mostly follows Kükenthal (1909)

Carex remota L.

Species	Section	Chromos. no. (2n)*
Carex repens Bell.	Arenariae Kunth	70
Carex siccata Dewey	Arenariae Kunth	70
Carex spicata Huds.	Muehlenbergianae Tuck.	58, 60
Carex stenophylla Wahlenb.	Physodeae Christ ex Kük.	60
Carex stipata Muehlenb.	Stenorhynchae Th. Holm	48, 52
Carex vallicola Dewey	Muehlenbergianae Tuck.	
Carex vernacula L. H. Bailey	Foetidae Tuck.	
Carex vulpina L.	Vulpinae (Carey) Christ	68
Carex vulpinoidea Michx.	Multiflorae Kunth	52, 54

Table 2 (continued)

* Chromosome counts compiled from: Böcher 1938; Dalgaard 1991; Davies 1956; Delay 1971; Dietrich 1972; Heilborn 1922, 1924, 1928, 1939; Kjellqvist and Löve 1963; Löve and Löve 1981; Moore and Calder 1964; Moore and Chater 1971; Murín and Májovsky 1976; Naczi 1999; Rothrock and Reznicek 1996; Tanaka 1942a, 1942b, 1948; Whitkus 1981, 1991 and fide FNA 2002

It is worth mentioning that Kükenthal (1909) arranged the monotypic section *Incurvae* at the beginning of the sections in subgenus *Vignea*, unfortunately without any explanation. The separation from section *Physodeae* and section *Foetidae* is limited to the contour of spikes and the number of spikelets.

In our trees, C. vallicola and C. macrocephala appear as members of subgenus Vignea, each in an isolated position. C. vallicola, distributed in pacific North America, is traditionally ascribed to section *Muehlenbergianae*. This heterogeneous section is divided into almost six different parts in our dendrograms. However, a core group can be detected and is discussed below. C. macrocephala is the only three-stigmatic species integrated in our analyses. The isolated position of this species supports the concept of section Macrocephalae (e.g. Kükenthal 1909, Mackenzie 1931-1935, Ohwi 1936, Egorova 1999, Mastrogiuseppe 2002). On the other hand it can be concluded that C. macrocephala and presumably also its East Asian sister taxon C. kobomugi Ohwi are true members of subgenus Vignea. These species can not be separated based on the presence of three stigmata as Kreczetovicz (1935) proposed by establishing the new subgenus Megalocranion Kreczetovicz.

Sections Chordorrhizeae, Heleonastes and Ovales. With the exception of *C. chordorrhiza*,

this group as a whole is characterized by inflorescences with gynaecandrous spikes.

Members of section Heleonastes cluster together, except C. canescens, which therefore is referred to as a single member of section Canescentes in our dendrograms. C. lachenalii, known from Europe and North America (as C. bipartita All.) occurs as sister taxon to Eurasian C. loliacea, both sharing subarctic distribution. Kükenthal (1909) ascribed C. loliacea to section Tenuiflorae, the sistersection of Canescentes. Later authors (e.g. Mackenzie 1931–1935, Kreczetovicz 1935, Hylander 1966) transferred this species to section Heleonastes in a broader sense, comprising sections Canescentes and Tenuiflorae. C. heleonastes, the name-giving species, is connected to them at short distance. Also C. brunnescens can be integrated into this highly supported group (a posteriori probability 100%, bootstrap value 91%). Surprisingly, the striking morphological similarity between C. brunnescens and C. canescens is not reflected by molecular data.

Carex chordorrhiza is characterized by a unique gross-morphology with long-creeping overground tillers and preference of very moist to wet habitats. It was ascribed to section *Divisae* by most caricologists (e.g. Kükenthal 1909, Mackenzie 1931–1935, Ohwi 1936, Chater 1980, Egorova 1999) and placed in section Acroarrhenae, subsection Foetidae by Bailey (1886). Fries (1845) established the section Chordorrhizeae, which was accepted by Mackenzie (1931–35) and Reznicek and Catling (2002) and is used in our dendrograms. The closer relationship to the core group of Heleonastes is well supported in both analyses (a posteriori probability 100%, bootstrap value 99%).

Species of section Ovales are distributed mainly in North and South America, from the Arctic to Patagonia, with the greatest diversity in the mountains of the western United States (Reznicek 1993). Kükenthal (1909) listed 21 species in this section, combining many similar taxa as subspecies and varieties under a comprehensive species name. Mackenzie (1931-35) favored a narrow species concept and listed 73 taxa for North America, Mastrogiuseppe et al. (2002) accepted 72 species. In contrast to other sections, the circumscription of section Ovales is rather precise: cespitose growth, spikelets gynaecandrous, perigynia flattened, more or less broadly winged. However, many transitions exist and species-delimitation in this section is therefore rather difficult (Reznicek 1993). Section Schellhammeria is separated from section Ovales only by the smaller perigynium. Therefore the position of C. bohemica within section Ovales could be expected and was shown by Roalson et al. (2001) in a smaller species sampling. Bailey (1886) and Mackenzie (1931-35) placed the American sister taxon C. synchnocephala Carey in section Ovales. The densely capitate inflorescence and the protruding leafy lowermost bracts of C. bohemica can also be found in species of section Ovales: C. athrostachya is characterized by a leafy basal bract.

The analyzed members of section Ovales form a very homogeneous group with high support in both dendrograms (*a posteriori* probability 100%, bootstrap value 94%). The name-giving Eurasian species C. ovalis is naturalized in North America and many other parts of the world. It clusters together with C. bicknellii and C. muskingumensis with high support. C. fracta and C. pinetorum appear closely related to this core group of Ovales in both dendrograms.

The South American species *C. bonplandii* with scarcely winged perigynium was treated as member of section *Elongatae* by Kükenthal (1909). It was referred to section *Ovales* by American authors (e.g. Bailey 1886, Mackenzie 1931–1935). *C. pinetorum* also has a weakly winged perigynium in comparison with *C. athrostachya*. Many transitions between wing-structures can be found in section *Ovales* (Reznicek 1993).

A specimen of *C. macloviana* from Swedish Lappland was studied. It clusters well in section *Ovales*, but the relationship to other species of the section remains unresolved. The disjunct distribution of *C. macloviana* and many closely related species in North America (Whitkus and Packer 1984, Whitkus 1988, Mastrogiuseppe et al. 2002) would be worth detailed investigations.

Sections *Elongatae*, *Muehlenbergianae* and *Vulpinae*. The molecularly highly supported cluster (*a posteriori* probability 100%, boot-strap value 98%) comprises species of different sections.

Carex elongata and *C. echinata* cluster together with high support, thus showing the difficult separation of section *Elongatae* from section *Muehlenbergianae*; the position of *C. remota* is discussed below. According to the molecular data, *C. elongata* is not closely related to section *Canescentes*, as was already assumed by Russian authors (Kreczetovicz 1935, Egorova 1966). The achene epidermis, densely covered with nodular protrusions of the many silica bodies (Toivonen and Timonen 1976), shows striking similarity between *C. elongata* and *C. echinata*. This morphology strongly supports the grouping based on molecular data.

Carex paniculata is widely distributed in the northern hemisphere, Australia and New Zealand. It is one of the conspicuous sedges growing in large tussocks. Although it appears to be very similar to *C. appropinquata* and *C. diandra* in gross morphology and in chromosome number, the three species, classically included in section *Paniculatae*, appear at separate positions in the molecular trees.

The core group of section Muehlenbergianae, although not highly supported (a posteriori probability 83%, bootstrap value 78%), comprises 4 very similar species mainly distributed in Europe. In many floras it is referred to as aggregate of C. muricata. C. divulsa was introduced in North America and other parts of the world. The distinction to C. leersii is not easy, therefore the latter is often treated as subspecies of C. divulsa (e.g. Chater 1980, Sebald 1998). According to ITS data, these two species are well separated. C. leersii, collected in France, seems closely related to C. muricata from Estonia. C. spicata, morphologically characterized by the swollen spongy basal part of the perigynium, is clearly separated in our dendrograms. It is distributed in Eurasia and North Africa and was also introduced to North America. To clarify the circumscription of the fairly diverse section *Muehlenbergianae* a larger sampling, especially of North American species, is required.

In both analyses, section *Vulpinae* is a sister group of the core cluster of section *Muehlenbergianae*. The two species studied, *C. vulpina*, the True fox sedge and *C. otrubae*, the False fox sedge, share identical ITS sequences. Thus, the morphologically closely related taxa cannot be distinguished by ITS data.

Carex canariensis, a local endemic of the Canary Islands described by Kükenthal (1900) and referred to section *Muehlenbergianae*, is a close relative to the group. Kükenthal (1900) postulated differences to *C. paniculata*, which is supported by ITS data.

Carex foetida, the name-giving species of section *Foetidae*, is distributed in the central and western Alps and the Pyrenees. It is adapted to moist meadows covered by snow for long periods. A basal position in subgenus *Vignea*, as was supposed by Kükenthal (1909), is not supported by molecular results.

The group including *C. canariensis*, *C. foetida* and the core-groups of sections *Muehlenbergianae* and *Vulpinae* is supported in

MCMC analysis with 100%, in NJ with only 62%. It is characterized by an almost stable chromosome number of 2n = 58. A single chromosome count of *C. vulpina* (2n = 68) has to be verified. Also species of section *Arenariae* form a highly supported lineage in molecular trees and share a stable chromosome number of 2n = 58.

Regarding the highly supported larger cluster (*a posteriori* probability 100%, boot-strap value 98%), including *C. paniculata* and section *Elongatae*, chromosome numbers are rather homogeneous reaching from 56 to 58, in *C. paniculata* up to 62.

Sections Deweyanae, Multiflorae, Remotae, Canescentes, Dioicae, Physodeae and Arenariae. The terminal group appears well supported in MCMC analysis. In NJ analysis C. canescens is placed distantly and the whole cluster is not supported. The unexpected position of C. canescens is discussed below.

Carex divisa and *C. diandra* cluster together with *C. bromoides* at a basal position in this terminal group. *C. divisa* seems closely related to *C. diandra* in MCMC analysis, whereas the NJ dendrogram reveals a closer relationship to *C. bromoides*, both positions without significant support.

In its gross morphology *C. diandra* is a slightly atypical member of section *Panicula-tae*, normally not growing in dense tussocks. The perigynium is significantly smaller than in other *Paniculatae*-species. There are also micromorphological differences in achene epidermis structures (Toivonen and Timonen 1976). In our molecular tree *C. paniculata* and *C. appropinquata* are separated and belong to different clades.

The next group is weakly supported in MCMC analysis (*a posteriori* probability 67%). It comprises mainly North American species of different traditional sections, indicating that the separation of sections *Muehlenbergianae*, *Stenorhynchae* and *Multiflorae* might be artificial.

Carex stipata is distributed in North America and East Asia and is treated in different sections in the regional floras (e.g. Kükenthal 1909, Mackenzie 1931–1935, Ohwi 1936). The only European species in this group is *C. appropinquata*. It is a typical member of section *Paniculatae*. Gross morphology and ultrastructure of achene epidermis (Toivonen and Timonen 1976) suggest a closer relationship to *C. paniculata*. However, this is not supported by ITS data. The three members of section *Paniculatae* integrated in our analyses do not appear closely related to each other; yet the unsupported positions in the ITS dendrograms do not allow conclusive groupings.

Carex densa and *C. vulpinoidea* cluster together, but they are clearly separated from *C. alma*, the third species of section *Multiflorae* in our analyses. *C. vulpinoidea* originated from temperate North America and is naturalized in parts of Europe.

Carex leavenworthii is one of nine species of the inhomogeneous section Muehlenbergianae studied in our analyses. Kükenthal (1909) separated section Muehlenbergianae from section Bracteosae on the basis of rhizome development and the basal structure of the perigynium. Mackenzie (1931-35) united both sections to one species-rich section Bracteosae. Ball (2002) considered section Bracteosae to be endemic to Central and South America and accepted sections Phaestoglochin and Stellulatae for North America, which were established by Egorova (1966, 1999) for the Flora of Russia. None of the proposed classifications corresponds with our present molecular results. Therefore only the core group of Muehlenbergianae is labeled in our dendrograms (see above).

The interpretation of *C. vernacula* as an American variety of the European *C. foetida* (e.g. Kükenthal 1909, Reznicek 2002) is not supported by ITS data. *C. vernacula* and *C. foetida* appear in distant positions in both dendrograms. The closer relationship between *C. vernacula* and *C. jonesii* is well supported in both analyses (*a posteriori* probability 100%, bootstrap value 81%).

Carex canescens is represented in our analyses by a Siberian and a French specimen

differing in only 1 bp in ITS sequence. The distant position of *C. canescens* to other members of section *Heleonastes* in molecular trees is surprising and was never recognized in other caricological studies. *C. canescens* is known to hybridize easily with species of nearly all sections in subgenus *Vignea* (Hylander 1966, Flatberg 1972, Toivonen 1981). This extreme hybridization potential is shared only by *C. remota* and *C. echinata* and is still not understood. Morphological structures of the perigynium and the achene epidermis, (Toivonen and Timonen 1976) cannot be used to explain the exceptional position of *C. canescens* revealed by ITS data either.

Kükenthal (1909) placed C. remota in section *Elongatae*, together with *C. elongata* and C. echinata, the latter erroneously named C. stellulata Good. (Kükenthal 1911). Microscopic structures of the achene epidermis in C. remota differ considerably from those of other members of section Elongatae (Toivonen and Timonen 1976), thus supporting our molecular result. C. remota is positioned differently in both analyses, but it has no close relationship to members of sections *Elongatae* and Canescentes, contrary to what had been postulated by Egorova (1966). We therefore separate sections Remotae and Elongatae in our dendrograms. C. remota is distributed from northern Africa to northern Europe up to 63° North. It is known as one of the most intensively hybridizing Carex species. C. remota hybridizes not only with C. arenaria, C. brizoides, and C. canescens, but also with C. appropinguata, C. otrubae, C. divulsa, C. spicata, C. paniculata, C. echinata, C. elongata, and C. ovalis (Kükenthal 1909, Nannfeldt 1977, Toivonen 1981, Jermy et al. 1982, Sebald 1998).

Carex alma, traditionally placed in section *Multiflorae*, and *C. macrorrhiza*, ascribed to section *Divisae*, cluster together with high support. In MCMC analysis these taxa occur in sister position to *C. canescens*.

Carex duriuscula was treated as the Siberian variety of *C. stenophylla* by Kükenthal (1909). In our analyses, these two species are

well separated by long branches, but show no closer relationship to any other member of section *Divisae*; therefore we labeled this well supported group as section *Physodeae* (Chater 1980, Egorova 1999). It occurs in sister position to the three European members of section *Dioicae*, *C. davalliana*, *C. dioica* and *C. parallela*.

Kükenthal (1909) placed section Dioicae in subgenus Primocarex. The vast majority of caricologists (e.g. Bailey 1889, Heilborn 1922, Kreczetovicz 1935, Ohwi 1936, Nelmes 1952, Egorova 1966, Toivonen and Timonen 1976, Nannfeldt 1977, Chater 1980, Cochrane 2002) included section Dioicae in subgenus Vignea, which is supported by our data. The terminal spike can consequently be interpreted as a reduced composite spike with only one-dimensional lateral branches. The common hybridization with species of subgenus Vignea (C. canescens, C. maritima, C. lachenalii, C. echinata) was already mentioned by Kükenthal (1909). Additional and independent support for the integration of section Dioicae in subgenus Vignea is lent through the relationship of Carex-specific parasitic smut fungi (e.g. Nannfeldt 1968, 1977). In MCMC analysis, C. dioica and C. parallela appear closely related, in the NJ tree a closer relationship of C. davalliana to C. parallela is indicated, as it is also by chromosome numbers (comp. Table 2).

The terminal cluster comprises species ascribed to section Arenariae with Eurasian distribution. This group includes C. arenaria as name-giving species and is supported in both analyses (a posteriori probability 100%, bootstrap value 82%). A heterogeneous assemblage of species, traditionally placed in Arenariae, was discussed above and is apparently not belonging to the core group as revealed by our data. We concentrate in the following on the well supported terminal Arenariae-group. Within this clade, only C. arenaria and C. ligerica appear closely related in both analyses. C. arenaria is well adapted to sandy habitats, distributed mainly in the coastal areas of temperate Europe. We studied a specimen from inland sands of eastern Germany. The group as a whole is also characterized by almost similar chromosome numbers of 2n = 58 (*C. arenaria* up to 64).

Carex praecox and C. brizoides are well characterized species, distributed in southern and continental Europe except the northern regions. Taxonomically C. curvata has been treated differently in European floras. Parent (1974), Chater (1980) and Sebald (1998) considered it as a common hybrid of C. praecox and C. brizoides. Kükenthal (1892, 1909) and Schultze-Motel (1967-1980) treated it as a subspecies of C. praecox. Molecular analyses reveal a closer relationship to C. praecox than to C. brizoides, without giving a clue to the origin of the taxon. C. pseudobrizoides was treated as 'species incertae sedis' by Kükenthal (1909). Chater (1980) presumed identity with C. reichenbachiana Bonnet; Luceño (1994) postulated identity with C. brizoides. The phylogenetic position of C. pseudobrizoides is not congruent in the two dendrograms; a close relationship with C. praecox, as suggested by MCMC analysis, is only weakly supported.

An interesting morphological character of the *Arenariae*-group is the existence of a scalelike prophyll at the basis of spikes. It appears to be absent in all other species of subgenus *Vignea* (Kükenthal 1909, Egorova 1999).

General aspects of the subgenus *Vignea*. Subgenus *Vignea* was considered to be a natural group within genus *Carex* by most caricologists. This morphological concept was recently confirmed by ITS and chloroplast DNA sequence data (Starr et al. 1999, Yen and Olmstead 2000, Roalson et al. 2001).

The delimitation of sections within this subgenus is very difficult and remains artificial in most parts. However, our molecular results allow better sectional circumscriptions and understanding for the species studied. Thus, the relationships between sections are well resolved by ITS sequences, revealing at least four larger subgroups, which are highly supported in all analyses (see above).

Chromosome numbers (Table 2) appear rather homogeneous in subgenus Vignea, not

indicating whether fission or fusion of polycentric chromosomes was predominant. It is noticeable that two groups revealed by molecular analyses might be characterized by their chromosome numbers. The well supported group including *C. canariensis*, *C. foetida* and the core groups of sections *Muehlenbergianae* and *Vulpinae* and also the terminal cluster including members of section *Arenariae* share a chromosome number of 2n = 58.

Although the composition of some sections has become clearer in the course of the present study, no taxonomic conclusions are drawn. As was argued for subgenus *Carex* (Hendrichs et al. 2004), a worldwide approach is necessary to understand the natural groups within the subgenus *Vignea*.

Even after 200 years of intensive work in the genus *Carex*, we are far from understanding the natural delimitations of sections within the well-defined subgenus *Vignea*. Due to the limited number of species studied so far answer only a few problems could be solved and many open questions remain. Nevertheless, the presented dendrograms can give at least new stimulation for detailed investigations in morphological characters of the species. Ultrastructural data of achene epidermis (Toivonen and Timonen 1976, Starr and Ford 2001) and microscopic features of leaf anatomy (Shepherd 1976) have shown some potential and their exploration should be resumed.

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References

Bailey L. H. (1886) A preliminary synopsis of North American *Carices*, including those of Mexico, Central America, and Greenland, with the American bibliography of the genus. Proc. Amer. Acad. Arts 22: 59–157.

- Bailey L. H. (1889) *Carex*. In: Gray A. (ed.) The manual of the botany of the Northern United States. American Book Co., New York, pp. 587–622.
- Ball P. W. (1990) Some aspects of the phytogeography of *Carex*. Canad. J. Bot. 68: 1462–1472.
- Ball P. W. (2002) Section *Phaestoglochin*. In: Flora of North America Editorial Committee (ed.) Flora of North America. Oxford University Press, New York, pp. 285–297.
- Ball P. W., Reznicek A. A. (2002) *Carex* Linnaeus. In: Flora of North America Editorial Committee (ed.) Flora of North America. Oxford University Press, New York, pp. 254–572.
- Böcher T. W. (1938) Zur Zytologie einiger arktischen und borealen Blütenpflanzen. Svensk Bot. Tidskr. 32: 346–361.
- Chater A. O. (1980) Carex L. In: Tutin T. G., Heywood V. H., Burges N. A., Valentine D. H., Walter S. M., Webb D. A. (eds.) Flora Europaea. Cambridge University Press, Cambridge, pp. 290–323.
- Cochrane T. S. (2002) Section *Physoglochin*. In: Flora of North America Editorial Committee (ed.) Flora of North America. Oxford University Press, New York, pp. 299–301.
- Dalgaard V. (1991) Chromosome studies in flowering plants from Macaronesia II. Willdenowia 20: 139–152.
- Davies E. W. (1956) Cytology, evolution and origin of the aneuploid series in the genus *Carex*. Hereditas 42: 349–365.
- Delay J. (1971) Halophytes et écotypes maritimes. Inf. Ann. Caryosyst. Cytogenet. 5: 29–40.
- Dietrich W. (1972) IOPB chromosome number reports. XXXVI. Taxon 21: 333–335.
- Drejer S. (1844) Symbolae Caricologicae ad synonymiam Caricum extricandam stabiliendamque et affinitates naturales eruendas. Halfniae, Bianco Luno.
- Dumortier B. C. J. (1827) Florula Belgica, operis majoris prodomus. Staminatia. Tornati Nerviorum/Tournay.
- Egorova T. V. (1966) Osoki SSSR, vidy podroda *Vignea* (Subgen. *Vignea* in the U.S.S.R.). Moskva & Leningrad, Akademija Nauk SSSR.
- Egorova T. V. (1999) The sedges (*Carex* L.) of Russia and adjacent states (within the limits of the former USSR). St. Petersburg, Russian Academy of Sciences, Komarov Botanical Institute.

- Flatberg K. J. (1972) Carex x lidii Flatb. = C. canescens L. x chordorrhiza Ehrh. Norweg. J. Bot. 19: 91–106.
- Fries E. M. (1835) Corpus florarum provincialium Sueciae. I. Uppsala, Palmblad, Sebel & Co.
- Hall T. A. (1999) BioEdit: a user-friendly biological sequence alignment editor and analysis program for Windows 95/98/NT. Version 5.0.9. Nucleic Acids Symp. Ser. 41: 95–98.
- Heilborn O. (1922) Die Chromosomenzahlen der Gattung Carex. Svensk Bot. Tidskr. 16: 271–274.
- Heilborn O. (1924) Chromosome numbers and dimensions, species-formation and phylogeny in the genus *Carex*. Hereditas 5: 129–216.
- Heilborn O. (1928) Chromosome studies in Cyperaceae. I-II. Hereditas 11: 182–192.
- Heilborn O. (1939) Chromosome studies in Cyperaceae III-IV. Hereditas 25: 224–240.
- Hendrichs M., Michalski S., Begerow D., Oberwinkler F., Hellwig H. (2003a) Phylogenetic relationships in *Carex* subgenus *Vignea* (Cyperaceae).
 16th Intern. Symposium of the German Bot. Society. Palm. Hortus Francof. 7: 165 (abstract).
- Hendrichs M., Begerow D., Oberwinkler F. (2003b) The genus *Carex* (Cyperaceae) A phylogenetic hypothesis. 16th Intern. Symposium of the German Bot. Society. Palm. Hortus Francof. 7: 44 (abstract).
- Hendrichs M., Oberwinkler F., Begerow D., Bauer R. (2004) *Carex*, subgenus *Carex* (Cyperaceae) – A phylogenetic approach using ITS sequences. Plant Syst. Evol. (in press).
- Hsiao C., Chatterton N. J., Asay K. H., Jensen K.
 B. (1995) Molecular phylogeny of the Pooideae (Poaceae) based on nuclear rDNA (ITS) sequences. Theor. Appl. Genet. 90: 389–398.
- Huelsenbeck J. P., Ronquist F. (2001) MrBayes: Bayesian inference of phylogenetic trees. Bioinformatics 17: 754–755.
- Hylander N. (1966) Nordisk kärlväxtflora. Stockholm, Almqvist & Wiksell.
- Jeanmougin F., Thompson J. D., Gouy M., Higgins D. G., Gibson T. J. (1998) Multiple sequence alignment with Clustal X. TIBS 23: 403–405.
- Jermy A. C., Chater A. O., David R. W. (1982) Sedges of the British Isles. London, Botanical Society of the British Isles.
- Kjellqvist E., Löve A. (1963) Chromosome numbers of some *Carex* species from Spain. Bot. Not. 116: 241–248.

- Kreczetovicz V. I. (1935) Carex L. In: Shishkin B. K., Komarov V. L. (eds.) Flora of the SSSR. Translated from Russian, Israel Program for Scientific Translation, Jerusalem 1964, Leningrad, pp. 111–464/86–369.
- Kükenthal G. (1909) Cyperaceae–Caricoideae. In: Engler A. (ed.) Das Pflanzenreich. Engelmann, Leipzig, pp. 1–824.
- Kükenthal G. (1911) Einiges über die Bearbeitung der Gattung *Carex* in J. Briquet, Prodrome de la Flore Corse. Allg. Bot. Z. Syst. 17: 150–151.
- Kunth C. S. (1837) Cyperographia synoptica sive enumeratio Cyperacearum omnium hucusque cognitarum, adjectis chracteribus, differentiis et synonymis. In: Kunth C. S. (ed.) Enumeratio plantarum omnium hucusque cognitarum, secudum familias naturalis disposita, adjectis chracteribus, differentiis et synonymis. J. G. Cotta, Stuttgart & Tübingen, pp. 368–592.
- Löve A., Löve D. (1981) Chromosome number reports. LXXIII. Taxon 30: 845–849.
- Mackenzie K. K. (1931–1935) Cyperaceae–Cariceae. The New York Botanical Garden, New York.
- Mastrogiuseppe J. (2002) Section *Macrocephalae*. In: Flora of North America Editorial Committee (ed.) Flora of North America. Oxford University Press, New York, pp. 307–309.
- Mastrogiuseppe J., Rothrock P. E., Dibble A. C., Reznicek A. A. (2002) Section *Ovales*. In: Flora of North America Editorial Committee (ed.) Flora of North America. Oxford University Press, New York, pp. 332–378.
- Moore D. M., Chater A. O. (1971) Studies of bipolar disjunct species I. *Carex*. Bot. Not. 124: 317–334.
- Moore R. J., Calder J. A. (1964) Some chromosome numbers of *Carex* species of Canada and Alaska. Canad. J. Bot. 42: 1387–1391.
- Muasya A. M., Simpson D. A., Chase M. W., Culham A. (1998) An assessment of suprageneric phylogeny in Cyperaceae using *rbcL* DNA sequences. Plant Syst. Evol. 211: 257–271.
- Murín A., Májovsky J. (1976) IOPB chromosome number reports. LIII. Taxon 25: 487.
- Naczi R. F. C. (1999) Chromosome numbers of some eastern North American species of *Carex* and *Eleocharis* (Cyperaceae). Contr. Univ. Michig. Herb. 22: 105–119.
- Nannfeldt J. A. (1968) Fungi as Plant Taxonomists. Acta Univ. Uppsal., 85–95.

- Nannfeldt J. A. (1977) The species of *Anthracoidea* (Ustilaginales) on *Carex* subgen. *Vignea* with special regard to the Nordic species. Bot. Not. 130: 351–375.
- Nelmes E. (1952) Facts and speculations on phylogeny in the tribe *Cariceae* of the Cyperaceae I. General considerations. Kew Bull. 1951: 427–436.
- Ohwi J. (1936) Cyperaceae Japonicae I. A synopsis of the *Caricoideae* of Japan, including the Kuriles, Saghalin, Korea, and Formosa. Memoirs of the College of Science / Kyoto University. Series B 11: 229–530.
- Posada D., Crandall K. A. (1998) Modeltest: testing the model of DNA substitution. Bioinformatics 14: 817–818.
- Rambaut A. (2001) Se-Al v2.0a72, Sequence Alignment Editor. Oxford UK, University of Oxford.
- Reznicek A. A. (1993) Revision of *Carex* section *Ovales* (Cyperaceae) in Mexico. Contr. Univ. Michig. Herb. 19: 97–136.
- Reznicek A. A. (2002) Section *Foetidae*. In: Flora of North America Editorial Committee (ed.) Flora of North America. Oxford University Press, New York, pp. 309–311.
- Reznicek A. A., Catling P. M. (2002) Section *Chordorrhizae*. In: Flora of North America Editorial Committee (ed.) Flora of North America. Oxford University Press, New York, pp. 298–299.
- Roalson E. H., Columbus J. T., Friar E. A. (2001) Phylogenetic relationships in *Cariceae* (Cyperaceae) based on ITS (nrDNA) and *trn*T-L-F (cpDNA) region sequences: Assessment of subgeneric and sectional relationships in *Carex* with emphasis on section *Acrocystis*. Syst. Bot. 26: 318–341.
- Rothrock P. E., Reznicek A. A. (1996) Documented chromosome numbers 1996: 1. Chromosome numbers in *Carex* section *Ovales* (Cyperaceae) from eastern North America. Sida Contrib. Bot.17: 251–258.
- Saitou N., Nei M. (1987) The neighbor-joining method: a new method for reconstructing phylogenetic trees. Mol. Biol. Evol. 4: 406–425.
- Schweinitz L. D. de (1825) Monograph of the North-American species of the genus *Carex*. Ann. Lyceum Nat. Hist. New York 1: 283–373.
- Sebald O. (1998) Carex L. 1753. In: Sebald O., Seybold S., Philippi G., Wörz A. (eds.) Die Farn- und Blütenpflanzen Baden-Württembergs. Ulmer, Stuttgart, pp. 98–248.

- Shepherd G. J. (1976) The use of anatomical characters in the intrageneric classification of *Carex* (Cyperaceae). Hoehnea 6: 33–54.
- Starr J. R., Bayer R. J., Ford B. A. (1999) The phylogenetic position of *Carex* section *Phyllo-stachys* and its implications for phylogeny and subgeneric circumscription in *Carex* (Cyperaceae). Amer. J. Bot. 86: 563–577.
- Starr J. R., Ford B. A. (2001) The taxonomic and phylogenetic utility of vegetative anatomy and fruit epidermal silica bodies in *Carex* section *Phyllostachys* (Cyperaceae). Canad. J. Bot. 79: 362–379.
- Swofford D. L., Olsen G. J., Waddell P. J., Hillis D. M. (1996) Phylogenetic Inference. In: Hillis D. M., Moritz C., Mable B. K. (eds.) Molecular systematics. Sinauer, Sunderland, pp. 407–514.
- Swofford D. L. (2002) PAUP*. Phylogenetic Analysis Using Parsimony (* and Other Methods). Version 4.0b10. Sunderland, Massachusetts, Sinauer Associates.
- Tanaka N. (1942a) Chromosome studies in Cyperaceae. XXI–XXIII. Chromosome numbers of *Eucarex* species (1–3). Med. & Biol., Tokyo 2: 289–300.
- Tanaka N. (1942b) Chromosome studies in Cyperaceae. XXIV–XXVI. Chromosome numbers of *Eucarex* species (4–6). Med. & Biol., Tokyo 2: 419–428.
- Tanaka N. (1948) The problem of aneuploidy. Biol. Contrib. in Japan 4: 1–327.
- Toivonen H. (1981) Spontaneous Carex hybrids of Heleonastes and related sections in Fennoscandia. Acta Bot. Fenn. 116: 1–51.
- Toivonen H., Timonen T. (1976) Perigynium and achene epidermis in some species of *Carex*, subg. *Vignea* (Cyperaceae), studied by scanning electron microscopy. Ann. Bot. Fenn. 13: 49–59.
- Tuckerman E. (1843) Enumeratio methodica caricum quarundam. Species recensuit et secundum habitum pro viribus disponere tentavit. Schenectady, NY State, USA, Isaac Riggs.
- Waterway M. J., Olmstead R. G. (1998) Phylogenetic relationships in *Carex* section *Hymenochlaenae* inferred from non-coding nuclear and chloroplast DNA sequence data (abstract). Amer. J. Bot. 85: 165.
- White T. J., Bruns T. D., Lee S., Taylor J. W. (1990) Amplification and direct sequencing of fungal ribosomal RNA genes for phylogenetics. In: Innis M. A., Gelfand D. H., Sninsky J. J.,

White T. J. (eds.) PCR protocols: A guide to methods and applications. Academic Press, Inc., San Diego, Calif, pp. 315–322.

- Whitkus R. (1981) Chromosome numbers of some northern New Jersey Carices. Rhodora 83: 461– 464.
- Whitkus R. (1988) Experimental hybridizations among chromosome races of *Carex pachystachya* and the related species *C. macloviana* and *C. preslii* (Cyperaceae). Syst. Bot. 13: 146–153.
- Whitkus R. (1991) Chromosome counts of *Carex* section *Ovales*. Bot. Gaz. (Crawfordsville) 152: 224–230.
- Whitkus R., Packer J. G. (1984) A contribution to the taxonomy of the *Carex macloviana* aggregate (Cyperaceae) in western Canada and Alaska. Canad. J. Bot. 62: 1592–1607.

Yen A. C., Olmstead R. G. (2000) Molecular systematics of Cyperaceae tribe *Cariceae* based on two chloroplast DNA regions: *ndh*F and *trn*L intron-intergenic spacer. Syst. Bot. 25: 479–494.

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